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TITLE OF INVENTION: Glucagon-Like Peptide-1 Improves the Ability of the B-Cell to Sense and Respond to Glucose in Subjects with Impaired Glucose Tolerance

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Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. [X] This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. [ ] This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. [X] This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. [ ] A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. [X] A copy of the International Application as filed (35 U.S.C. 371(c)(2))
   a. [ ] is transmitted herewith (required only if not transmitted by the International Bureau).
   b. [ ] has been transmitted by the International Bureau.
   c. [X] is not required, as the application was filed in the United States Receiving Office (RO/US).
6. [ ] A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. [X] Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
   a. [ ] are transmitted herewith (required only if not transmitted by the International Bureau).
   b. [ ] have been transmitted by the International Bureau.
   c. [ ] have not been made; however, the time limit for making such amendments has NOT expired.
   d. [X] have not been made and will not be made.
8. [ ] A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. [ ] An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. [ ] A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. [ ] An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. [ ] An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. [ ] A FIRST preliminary amendment.
    [ ] A SECOND or SUBSEQUENT preliminary amendment.
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GLUCAGON-LIKE PEPTIDE-1 IMPROVES β-CELL RESPONSE TO GLUCOSE IN SUBJECTS WITH IMPAIRED GLUCOSE TOLERANCE

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation-in-part of and claims the benefit of the filing date of U.S. Provisional Application No. 60/089,044 filed June 12, 1998.

BACKGROUND OF THE INVENTION

Impaired glucose tolerance (IGT) is common in the U.S. population. The prevalence of impaired glucose tolerance increases from 11% in the general population aged 20-74 years to 24% in those 40-75 years of age with a family history of diabetes and a body weight greater than 120% of normal. Subjects with impaired glucose tolerance are at high risk for the development of cardiovascular disease as well as non-insulin dependent diabetes mellitus (NIDDM), also known as Type 2 diabetes.

Impaired glucose tolerance is characterized by early subtle defects in pancreatic β-cell function, accompanied by insulin resistance. These early defects include an impaired ability of the β-cell to sense and respond to small changes in plasma glucose concentrations with appropriate levels of insulin secretion, and a mild shift to the right of the glucose insulin secretion dose-response curve. The glucose sensing and fast insulin secretion response abilities of the β-cell are lost very early in the course of IGT when 2-hour glucose levels are minimally elevated. The deterioration of glucose control in IGT with time is predominantly due to progressive impairment of β-cell function. This leads, in many cases, to deteriorating conditions of

hyperinsulinemia, obesity, and cardiovascular disease, sometimes known as Syndrome X. In many cases advanced IGT conditions lead to definitive loss of glucose control and the deleterious onset of NIDDM.

As indicated, the condition of IGT carries serious health risks. The IGT patient is often obese and has high plasma levels of insulin, which are often toxic. These high insulin levels result generally from the continually increased inability of muscle, other tissue and fat cells to utilize insulin to effect the uptake of glucose from blood plasma. The IGT condition gives rise to increased risks for a whole range of cardiovascular diseases.

Glucagon-like peptide-1 (GLP-1), a natural enteric peptide, is secreted from the L-cells of the gut, and acts as an incretin hormone stimulating pancreatic β-cells to secrete insulin in a glucose dependent manner. Its therapeutic potential in NIDDM has been previously demonstrated, in that exogenous infusion of pharmacological doses of GLP-1 generally reduced plasma glucose levels. However, GLP-1 did not significantly improve β-cell function in NIDDM. Nathan DM, Schreiber E, Fogel H, Mojsov S, Habener JF. Insulinotropic action of glucagon-like peptide-1 (7-37) in diabetic and nondiabetic subjects. *Diabetes Care* 15:270-276, 1992; Gutniak M, Ørskov C, Holst JJ, Ahrén B, Efendric S. Antidiabetogenic effects of glucagon-like peptide-1 (7-36) amide in normal subjects and patients with diabetes mellitus. *N Engl J Med* 326:1316-1322, 1992; Nauck MA, Kleine N. Orskov C, Holst JJ, Willms B, Creutzfeldt W. Normalization of fasting hyperglycemia by exogenous glucagon-like peptide-1 (7-36 amide) in type II (non-insulin-dependent) diabetic patients. *Diabetologia* 36:741-744, 1993; Gutniak MK, Linde B, Holst JJ, Efendie S. Subcutaneous injection of the incretin hormone glucagon-like peptide-1 abolishes postprandial glycemia NIDDM. *Diabetes Care* 17:1039-1044, 1994; Rachman J, Gribble FM, Barrow BA, Levy JC, Buchanan KD, Turner RC. Normalization of insulin responses to glucose by overnight infusion of glucagon-like peptide 1 (7-36) amide in patients with NIDDM. *Diabetes* 45:1524-1530, 1996; Rachman J, Barrow BA, Levy JC, Turner RC. Near-normalization of diurnal glucose concentrations by

continuous administration of glucagon-like peptide-1 (GLP-1) in subjects with NIDDM, Diabetologia 40:205-211, 1997.

The IGT condition is not currently treatable. It is, however, a recognizable disease condition associated with serious health risks. In general, the IGT condition progressively deteriorates in terms of its symptoms and often leads to loss of plasma glucose control which constitutes type 2 diabetes. There is a need for a therapy.

Numerous studies over the past several years have demonstrated that the application of GLP-1 in cases of NIDDM lowers glucose and insulin levels in the blood, and hence should be a promising therapy for that disease. However, no studies to date have shown that GLP-1 has a potential to correct the loss of the ability of β-cells to sense and quickly respond with the secretion of insulin when blood glucose increases. It is this deterioration in the ability to respond and to closely link the sensing of increases in blood glucose to insulin secretion from the β-cells which is the principal cause of the IGT condition. In previous studies the application of GLP-1 to NIDDM subjects demonstrated an ability to normalize fasting plasma glucose and to stimulate cumulative β-cell insulin secretion. However, GLP-1 infused overnight in NIDDM subjects did not improve glucose responses to meals the next day. When GLP-1 was infused for 19 hours, overnight and during three standard meals in subjects with NIDDM, plasma glucose levels were reduced, but the impaired post-prandial β-cell function was only slightly improved.

β-cell responses to prolonged infusion of GLP-1 have not been previously studied in subjects with IGT and, while there has been no indication that the result would be different than with GLP-1 infusions in NIDDMs, detailed studies of the effect of GLP-1 on β-cell responses to small increases and decreases in plasma glucose concentrations have not been heretofore performed.

Accordingly, a method to arrest the progression of IGT and restore normal glucose metabolism conditions is needed.

It is therefore an object of the present invention to provide a method of restoring or improving β-cell function and sensitivity, and thus insulin secretion patterns, in response to plasma glucose levels in a host having impaired glucose tolerance.

A further object of the invention is to provide a method to delay or prevent the deterioration of β-cell function which is responsible for the progression of impaired glucose tolerance into the loss of control over plasma glucose which characterizes the onset of NIDDM.

A yet further object of the invention is to ameliorate cardiovascular disease effects of IGT, thereby decreasing cardiovascular and stroke risks.

The method of accomplishing these and other objects will be apparent from the following detailed description.

SUMMARY OF THE INVENTION

The inventors have discovered that the application of GLP-1 in subjects with impaired glucose tolerance reestablishes the tightly coordinated response of insulin secretion to increases in plasma glucose levels, thereby restoring the insulin secretion response patterns from the β-cell to plasma glucose level increases which are characteristic of normal subjects without IGT.

Thus the present invention is directed to a method of treating a host having impaired glucose tolerance and insulin resistance with GLP-1 in an amount effective to restore, improve or normalize β-cell sensitivity and function and insulin secretion patterns in that host. The invention is also directed to a method of reducing plasma insulin level in persons with IGT and concurrently reducing the condition of insulin resistance and its concomitant condition of cardiovascular disease.

In carrying out the examples described herein, the inventors have surprisingly observed that the application of GLP-1 in subjects with IGT, in contrast to random and uncoordinated insulin response characteristically encountered in IGT subjects, dramatically recreates sensitive, quick and coordinated insulin secretions from the β-cells in response to discreet pulsatile

increases in plasma glucose similar to insulin secretion patterns found in normal patients.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the glucose, insulin and GLP-1 responses to oral administration of 75 mg of glucose in five subjects with impaired glucose tolerance (IGT •) and five subjects with non-insulin dependent diabetes mellitus (NIDDM □). Figure 1A shows mean glucose response. Figure 1B shows insulin response. Figure 1C shows GLP-1 response.

Figure 2 provides a comparison of mean insulin secretion rates (ISR) and mean glucose concentrations in each subject during glucose infusion with saline infusion (O) or GLP-1 infusion (•). Figure 2A shows the comparison in subjects with IGT. Figure 2B shows the comparison in subjects with NIDDM.

Figure 3 provides profiles of the glucose, insulin secretion rates (ISR) and insulin concentrations in two subjects with IGT, subjects D01 and D02. Figures 3A and 3C show the responses to saline infusion. Figures 3B and 3D show the responses to GLP-1 infusion.

Figure 4 provides a comparison of glucose levels, insulin secretion rates (ISR) and insulin levels in two subjects with NIDDM, subjects D07 and D09. Figures 4A and 4C show the profiles during saline infusion. Figures 4B and 4D show the profiles during GLP-1 infusion.

Figure 5 provides a comparison of spectral analyses of insulin secretion in subjects during saline and GLP-1 infusions. Results shown on the left side of the spectral analysis are of subjects with IGT. Results shown on the right side of the spectral analysis are of subjects with NIDDM.

Figure 6 provides a comparison of normalized spectral power during saline infusion and during GLP-1 infusion. Figure 6A shows a comparison of normalized spectral power during saline infusion and GLP-1 infusion in a subject with IGT (D02). Figure 6B shows a comparison of normalized spectral power during saline infusion and GLP-1 infusion in a subject with NIDDM (D07).

DETAILED DESCRIPTION OF THE INVENTION

The inventors have discovered that the administration of GLP-1 in subjects with impaired glucose tolerance (IGT) restored or improved pancreatic β-cell function and the ability of β-cells to respond quickly in secreting insulin in a coordinated manner in response to small increases or changes in plasma glucose concentrations, i.e. pulsatile secretions of insulin, similar to insulin secretion patterns found in subjects without IGT. This pattern of insulin secretion is not restored in subjects who have already developed NIDDM, which is characterized by loss of plasma glucose control.

β-cell function is quantified by normalized spectral power. Spectral power measures β-cell function which does not rely on adjustment for insulin sensitivity. The inventors have found that in subjects with IGT, GLP-1 improves spectral power into a normal range. The spectral power profiles indicated that the entrainment or close coordination of plasma glucose and insulin secretion oscillations was restored to normal levels in IGT subjects after administration of GLP-1. This improvement in the oscillatory pattern of insulin secretion is important for the maintenance of normal glucose homeostasis. For example, it has been shown that insulin infusions that mimic the ultradian oscillations within a period of 120 minutes are more effective than constant infusions of insulin in the reduction of plasma glucose concentrations (27).

The present invention provides a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and is effective in improving the ability of β-cells to sense and respond to small changes in plasma glucose concentrations in subjects with IGT. In one embodiment, the receptor-binding compound is glucagon-like peptide-1. In another embodiment, the receptor-binding compound is a variant peptide in which the combination of the substitutions, deletions and variants does not differ by more than ten amino acids from glucagon-like peptide-1. The receptor-binding compound can further comprise a polynucleotide or an agent which activates the release of GLP-1, a molecule which activates the GLP-1 receptor, or a GLP-

1 receptor binding compound comprising a chemically constructed molecule, peptide analogs, or agonists of GLP-1.

The inventors have discovered that the administration of human GLP-1 enhanced or restored entrainment of insulin secretion responses to small changes or increases in plasma glucose. Accordingly, the composition of the present invention is useful in therapeutic treatment for normalizing impaired glucose tolerance.

The inventors have demonstrated herein that a low dose infusion of GLP-1 can improve the function of the β-cells to secrete insulin in response to increases in plasma glucose levels. Thus, GLP-1 can also be used to improve the preservation of β-cell function in subjects with IGT. Administration of GLP-1 also regulates or normalizes insulin secretion patterns which will result in overall reduction of plasma insulin in IGT. This normalization in turn will reduce the condition of insulin resistance.

The term "GLP-1", or glucagon-like peptide, includes GLP-1 mimetics, and as used in the context of the present invention can be comprised of glucagon-like peptides and related peptides and analogs of glucagon-like peptide-1 that bind to a glucagon-like peptide-1 (GLP-1) receptor protein such as the GLP-1 (7-36) amide receptor protein and has a corresponding biological effect on insulin secretion as GLP-1 (7-36) amide, which is a native, biologically active form of GLP-1. See Göke, B and Byrne, M, *Diabetic Medicine*. 1996, 13:854-860. The GLP-1 receptors are cell-surface proteins found, for example, on insulin-producing pancreatic β-cells. Glucagon-like peptides and analogs will include species having insulinotropic activity and that are agonists of, i.e. activate, the GLP-1 receptor molecule and its second messenger activity on, *inter alia*, insulin producing β-cells. Agonists of glucagon-like peptide that exhibit activity through this receptor have been described: EP 0708179A2; Hjorth, S.A. et al., *J. Biol. Chem.* 269 (48):30121-30124 (1994); Siegel, E.G. et al. Amer. Diabetes Assoc. 57th Scientific Sessions, Boston (1997); Hareter, A. et al. Amer. Diabetes Assoc. 57th Scientific Sessions, Boston (1997); Adelhorst, K. et al. *J. Biol. Chem.*

269(9):6275-6278 (1994); Deacon C.F. et al. 16th International Diabetes Federation Congress Abstracts, *Diabetologia Supplement* (1997); Irwin, D.M. et al., *Proc. Natl. Acad. Sci. USA.* 94:7915-7920 (1997); Mosjov, S. *Int. J. Peptide Protein Res.* 40:333-343 (1992). Glucagon-like molecules include polynucleotides that express agonists of GLP-1, i.e. activators of the GLP-1 receptor molecule and its secondary messenger activity found on, *inter alia,* insulin-producing β-cells. GLP-1 mimetics that also are agonists of β-cells include, for example, chemical compounds specifically designed to activate the GLP-1 receptor. Glucagon-like peptide-1 antagonists are also known, for example see e.g. Watanabe, Y. et al., *J. Endocrinol.* 140(1):45-52 (1994), and include exendin (9-39) amine, an exendin analog, which is a potent antagonist of GLP-1 receptors (see, e.g. WO97/46584). Recent publications disclose Black Widow GLP-1 and Ser^2 GLP-1, see G.G. Holz, J.F. Hakner/*Comparative Biochemistry and Physiology*, Part B 121(1998)177-184 and Ritzel, et al., *A synthetic glucagon-like peptide-1 analog with improved plasma stability*, J. Endocrinol 1998 Oct.; 159(1):93-102.

Further embodiments include chemically synthesized glucagon-like polypeptides as well as any polypeptides or fragments thereof which are substantially homologous. "Substantially homologous," which can refer both to nucleic acid and amino acid sequences, means that a particular subject sequence, for example, a mutant sequence, varies from a reference sequence by one or more substitutions, deletions, or additions, the net effect of which does not result in an adverse functional dissimilarity between reference and subject sequences. For purposes of the present invention, sequences having greater than 50 percent homology, and preferably greater than 90 percent homology, equivalent biological activity in enhancing β-cell responses to plasma glucose levels, and equivalent expression characteristics are considered substantially homologous. For purposes of determining homology, truncation of the mature sequence should be disregarded. Sequences having lesser degrees of homology, comparable bioactivity, and equivalent expression characteristics are considered equivalents.

Mammalian GLP peptides and glucagon are encoded by the same gene. In the ileum the phenotype is processed into two major classes of GLP peptide hormones, namely GLP-1 and GLP-2. There are four GLP-1 related peptides known which are processed from the phenotypic peptides. GLP-1 (1-37) has the sequence His Asp Glu Phe Glu Arg His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:1). GLP-1 (1-37) is amidated by post-translational processing to yield GLP-1 (1-36) NH_2 which has the sequence His Asp Glu Phe Glu Arg His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:2); or is enzymatically processed to yield GLP-1 (7-37) which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:3). GLP-1 (7-37) can also be amidated to yield GLP-1 (7-36) amide which is the natural form of the GLP-1 molecule, and which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ.ID NO:4) and in the natural form of the GLP-1 molecule.

Intestinal L cells secrete GLP-1 (7-37) (SEQ. ID NO:3) and GLP-1(7-36)NH_2 (SEQ.ID NO:4) in a ratio of 1 to 5, respectively. These truncated forms of GLP-1 have short half-lives *in situ*, i.e., less than 10 minutes, and are inactivated by an aminodipeptidase IV to yield Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:5); and Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:6), respectively. The peptides Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:5) and Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:6), have been speculated to affect hepatic glucose

production, but do not stimulate the production or release of insulin from the pancreas.

There are six peptides in Gila monster venoms that are homologous to GLP-1. Their sequences are compared to the sequence of GLP-1 in Table 1.

TABLE 1

a. H A E G T F T S D V S S Y L E G Q A A K E F I A W L V K G R NH_2

b. H S D G T F T S D L S K Q M E E E A V R L F I E W L K N G G P S S G A P P P S NH_2

c. D L S K Q M E E E A V R L F I E W L K N G G P S S G A P P P S NH_2

d. H G E G T F T S D L S K Q M E E E A V R L F I E W L K N G G P S S G A P P P S NH_2

e. H S D A T F T A E Y S K L L A K L A L Q K Y L E S I L G S S T S P R P P S S

f. H S D A T F T A E Y S K L L A K L A L Q K Y L E S I L G S S T S P R P P S

g. H S D A I F T E E Y S K L L A K L A L Q K Y L A S I L G S R T S P P P NH_2

h. H S D A I F T Q Q Y S K L L A K L A L Q K Y L A S I L G S R T S P P P NH_2

a=GLP-1(SEQ. ID NO:4).

b=Exendin 3(SEQ. ID NO:7).

c=Exendin 4(9-39(NH_2(SEQ.ID NO:8).

d=Exendin 4(SEQ.ID NO:9).

e=Helospectin I(SEQ.ID NO:10).

f=Helospectin II(SEQ. ID NO:11).

g=Helodermin(SEQ. ID NO:12).

h=Q^8, Q^9 Helodermin(SEQ. ID No:13).

The major homologies as indicated by the outlined areas in Table 1 are: peptides c and h are derived from b and g, respectively. All 6 naturally

occurring peptides (a, b, d, e, f and g) are homologous in positions 1, 7, 11 and 18. GLP-1 and exendins 3 and 4 (a, b and d) are further homologous in positions 4, 5, 6, 8, 9, 15, 22, 23, 25, 26 and 29. In position 2, A, S and G are structurally similar. In position 3, residues D arid E (Asp and Glu) are structurally similar. In positions 22 and 23 F (Phe) and I (Ile) are structurally similar to Y (Tyr) and L (Leu), respectively. Likewise, in position 26 L and I are structurally equivalent.

Thus, of the 30 residues of GLP-1, exendins 3 and 4 are identical in 15 positions and equivalent in 5 additional positions. The only positions where radical structural changes are evident are at residues 16, 17, 19, 21, 24, 27, 28 and 30. Exendins also have 9 extra residues at the carboxyl terminus.

The GLP-1 like peptides can be made by solid state chemical peptide synthesis. GLP-1 can also be made by conventional recombinant techniques using standard procedures described in, for example, Sambrook and Maniaitis. "Recombinant", as used herein, means that a protein is derived from recombinant (e.g., microbial or mammalian) expression systems which have been genetically modified to contain an expression gene for GLP-1 or its biologically active analogues.

The GLP-1 like peptides can be recovered and purified from recombinant cell cultures by methods including, but not limited to, ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. High performance liquid chromatography (HPLC) can be employed for final purification steps.

The polypeptides of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from prokaryotic or eukaryotic hosts (for example by bacteria, yeast, higher plant, insect and mammalian cells in culture or *in vivo*). Depending on the host employed in a recombinant production procedure, the

polypeptides of the present invention are generally non-glycosylated, but may be glycosylated.

GLP-1 activity can be determined by standard methods, in general, by receptor-binding activity screening procedures which involve providing appropriate cells that express the GLP-1 receptor on their surface, for example, insulinoma cell lines such as RINmSF cells or INS-1 cells. See also Mosjov, S.(1992) and EP0708170A2. In addition to measuring specific binding of tracer to membrane using radioimmunoassay methods, cAMP activity or glucose dependent insulin production can also be measured. In one method, a polynucleotide encoding the receptor of the present invention is employed to transfect cells to thereby express the GLP-1 receptor protein. Thus, for example, these methods may be employed for screening for a receptor agonist by contacting such cells with compounds to be screened and determining whether such compounds generate a signal, i.e. activate the receptor.

Polyclonal and monoclonal antibodies can be utilized to detect purify and identify GLP-1 like peptides for use in the methods described herein. Antibodies such as ABGA1178 detect intact unspliced GLP-1 (1-37) or N-terminally-truncated GLP-1 (7-37) or (7-36) amide. Other antibodies detect on the very end of the C-terminus of the precursor molecule, a procedure which allows by subtraction to calculate the amount of biologically active truncated peptide, i.e. GLP-1 (7-37) or (7-36) amide (Orskov et al. Diabetes, 1993, 42:658-661; Orskov et al. *J. Clin. Invest.* 1991, 87:415-423).

Other screening techniques include the use of cells which express the GLP-1 receptor, for example, transfected CHO cells, in a system which measures extracellular pH or ionic changes caused by receptor activation. For example, potential agonists may be contacted with a cell which expresses the GLP-1 protein receptor and a second messenger response, e.g. signal transduction or ionic or pH changes, may be measured to determine whether the potential agonist is effective.

The glucagon-like peptide-1 receptor binding proteins of the present invention may be used in combination with a suitable pharmaceutical carrier.

Such compositions comprise a therapeutically effective amount of the polypeptide, and a pharmaceutically acceptable carrier or excipient. Such a carrier includes, but is not limited, to saline, buffered saline, dextrose, water, glycerol, ethanol, lactose, phosphate, mannitol, arginine, trehalose and combinations thereof. The formulations should suit the mode of administration and are readily ascertained by those of skill in the art. The GLP-1 peptide may also be used in combination with agents known in the art that enhance the half-life *in vivo* of the peptide in order to enhance or prolong the biological activity of the peptide. For example, a molecule or chemical moiety may be covalently linked to the composition of the present invention before administration thereof. Alternatively, the enhancing agent may be administered concurrently with the composition. Still further, the agent may comprise a molecule that is known to inhibit the enzymatic degradation of GLP-1 like peptides may be administered concurrently with or after administration of the GLP-1 peptide composition. Such a molecule may be administered, for example, orally or by injection.

GLP-1 can be administered intravenously or subcutaneously, and can be administered continuously or by bolus injection. Total administration can be together with, before or after glucose injection or infusion. The following doses can be used: For continuous infusion by intravenous (I.V.) 0.1 pmol/kg/min to 10 pmol/kg/min and by subcutaneous (S.C.) 0.1 pmol/kg/min to 75 pmol/kg/min, and for single injection (bolus) by I.V. 0.005 nmol/kg to 20 nmol/kg and S.C. 0.1 nmol/kg to 100 nmol/kg.

The preferred method of administration of the GLP-1 peptide is through a continuous application. However, GLP-1 can be delivered by subcutaneous, intramuscular, interperitoneal, injected depot with sustained release, deep lung insufflation with sustained release as well as by intravenous, buccal, patch or other delivery method.

The effective treatment of IGT also decreases the risk of cardiovascular and cerebrovascular events. It can therefore be provided as a preventative to patients of known high risk for such events.

The following examples further illustrate an aspect of the present invention. However, these examples should in no way be taken as a limitation of the teachings or disclosure of the present invention to the exemplary limits in terms of scope.

EXAMPLES

The studies described herein were performed in 10 subjects who were divided into two groups on the basis of their plasma glucose response to an oral glucose tolerance test using the criteria of the World Health Organization (21) to define the degree of glucose intolerance. Five subjects had IGT, and five subjects had NIDDM. The gender, age, body mass index (BMK), basal levels of fasting glucose, 2 hour glucose, fasting insulin, and HBA1c for each subject are presented in Table 2. Diabetic subjects were older than those with IGT, but the groups were matched for by BMI. Mean fasting glucose levels and glycosylated hemoglobin concentrations were lower in the IGT group compared to subjects with NIDDM. Fasting insulin levels did not differ between the groups.

Table 2

Baseline clinical parameters of IGT and NIDDM subjects

ID	Sex	Age	BMI	Fasting glucose (mM)	2 hr glucose (mM)	Fasting Insulin (pmol/L)	Glyco-hemoglobin
IGT							
D01	M	50	25.7	5.78	8.99	54.84	5.8
D02	F	52	26.8	5.94	10.52	79.80	5.7
D03	M	49	32.2	5.73	9.45	73.68	6.3
D04	M	42	30.6	5.99	9.89	35.52	5.9
D05	M	46	38.2	6.14	11.06	92.88	6.5
Mean±SE		47.8±1.7	30.7±2.2	5.92±0.07	9.98±0.37	67.3±10.0	6.04±0.15
NIDDM							
D06	M	53	26.4	8.81	16.27	30.78	6.7
D07	M	61	27.9	6.87	11.28	81.60	7.2
D08	M	60	34.2	7.66	15.05	37.44	5.9
D09	M	53	27.8	6.86	18.66	47.88	7.7
D10	M	66	23.9	8.34	12.9	24.84	8.1
Mean±SE		58.6±2.5	28.1±1.7	7.71±0.39	14.83±1.29	44.5±10.0	7.12±0.39
P value		P<0.009	P=0.36	P<0.002	P<0.007	P=0.15	P<0.04

Plasma glucose levels were measured by the glucose oxidase technique (YSI, 1500 G, Schlag Company, Bergisch-Gladbach, Germany). The coefficient of variation of this method was <2%. Plasma insulin was measured by the Abbott IMx Microparticle Enzyme Immunoassay. The average intraassay coefficient of variation was 5%. Plasma C-peptide was measured as previously described in (22), Faber OK, Binder C, Markussen, J, Heding LG, Naithani VK, Kuzuya H, Blix P, Horwitz DL, Rubenstein AH. Characterization of seven c-peptide antisera. *Diabetes* 27, Suppl 1:170-177, 1978. The lower limit of sensitivity of the assay was 0.02 pmol/ml and the intraassay coefficient of variation averaged 6%. Glucagon was measured by using a commercially available radioimmunoassay kit (Biermann, Bad Nauheim, Germany) and the

intraassay coefficient of variation averaged 8%. IR-GLP-1 was measured using the specific polyclonal antibody GA 1178 (Affinity Research, Nottingham, UK) (23). It has 100% reactivity with GLP-1 (1-36) amide and the truncated GLP-1 (7-36) amide. Immunoreactive GLP-1 like material was extracted from plasma samples on C-18 cartridges employing acetonitrile for elution of the samples. The detection limit of the assay was 2 fmol/tube. The antiserum did not crossreact with GIP, pancreatic glucagon, glicentin, oxyntomodulin or GLP-2. Intra- and interassay coefficients of variation were 3.4% and 10.4%, respectively.

All results are expressed as mean ± SEM. Data analysis was performed using the Statistical Analysis System (SAS Version 6 Edition, for Personal Computers, SAS Institute, Inc., Cary, NC). The significance of differences within the individual induced by GLP-1 infusion was determined using paired t tests. Differences were considered to be significant if $P<0.05$.

Standard kinetic parameters for C-peptide clearance adjusted for age, sex and body surface area were utilized (24), Van Cauter E, Mestrez, F, Sturis J, Polonsky KS. Estimation of insulin secretion rates from C-peptide levels: comparison of individual and standard kinetic parameters for C-peptide clearance. *Diabetes* 41:368-377, 1992. These parameters were used to derive, in each 15 minute interval between blood sampling, the ISR from the plasma C-peptide concentrations by deconvolution as previously described (25,26).

The diabetic subjects were treated with diet alone with the exception of one subject D07, who was previously treated with an oral hypoglycemic agent that was discontinued 4 weeks prior to the study. None of the diabetic patients had ever received insulin. All subjects were placed on a weight maintenance diet containing at least 200g of carbohydrate per day for two weeks before the study.

Each subject was studied on three separate occasions. All studies were performed after a 12-hour overnight fast beginning at 0700 unless otherwise stated with subjects in the recumbent position. An intravenous catheter was placed in each forearm, one for blood sampling and one for administration of

glucose and GLP-1 as needed. In all experiments, the arm containing the sampling catheter was maintained in a heating blanket to ensure arterialization of the venous sample. In the following examples, the GLP-1 was administered to the subjects in the GLP-1 (7-36) amide form.

EXAMPLE 2 (Oral glucose tolerance test)

Blood samples were drawn for the measurement of glucose, C-peptide, insulin, glucagon and GLP-1 at 30 minute intervals for 120 minutes after ingestion of 75 g glucose (Boehringer, Mannheim, Mannheim, Germany). Incremental areas under the curve (AUC) from 0 to 120 minutes were calculated for glucose, insulin, C-peptide, glucagon and GLP-1. The glucose concentrations were used to define the degree of glucose intolerance according to the criteria of the World Health Organization. The response of the IGT and NIDDM groups to oral glucose is set forth in Table 3.

The glucose, insulin and GLP-1 responses of subjects to 75 mg of glucose is shown in Figure 1. The AUC for glucose from 0 to 120 minutes was lower in the IGT group but the AUC for insulin, C-peptide, glucagon, and GLP-1 did not differ.

TABLE 3

Response to oral glucose

2hr AUC	IGT n=5	NIDDM n=5
Glucose (mM. min/l)	1290±41	1628±76*
Insulin (pmol. min/L)	53,750±10,648	26,083±10,047
C-peptide (pmol. min/L)	293±40	180±48
Glucagon (ng. min/L)	8130±1324	6858±920
GLP-1 (pmol. min/L)	805±141	983±111

*P<0.05 for IGT vs NIDDM

When mean ISRs were plotted against mean glucose levels it was observed that GLP-1 caused a significant reduction in glucose levels without significantly altering the mean insulin secretion rates (Figure 2).

EXAMPLE 3 (Administration of an oscillatory glucose infusion)

The peripheral administration of glucose in an oscillatory pattern results in regular oscillations in plasma glucose. In normal subjects, the β-cell is able to detect and respond to repetitive increases and decreases in glucose with parallel changes in insulin secretions. This adjustment of the insulin secretory oscillations to the glucose oscillations is termed entrainment. Lack of entrainment to glucose is an early manifestation of β-cell dysfunction in individuals with IGT and mild NIDDM.

We used a low-dose oscillatory glucose infusion protocol since it is a very sensitive test of the ability of the β-cell to sense and respond to small changes in plasma glucose concentrations. It tests the integrity of the feedback loop linking glucose and insulin secretion. A normal response requires an intact glucose sensing ability.

In order to determine whether the β-cells were able to detect and respond to oscillations of glucose, glucose was infused in an oscillatory pattern with a small volume of saline for 12 hours. The amplitude of the administered oscillations was 33% above and below the mean rate of 4 mg/kg/min and their periodicity was 144 minutes.

In order to establish the effect of GLP-1 on the ability of the β-cell to respond to oscillations of glucose, glucose was infused in the same manner and GLP-1 was infused at a constant rate of 0.4 pmol/kg/min for the entire 12 hours. Each study consisted of an initial 2-hour period (0700-0900) to allow a steady state to be achieved. This was followed by a subsequent period of 10 hours (0900-1900) during which time blood samples were drawn at 15 minute intervals for glucose, insulin, C-peptide, and glucagon and at 60 minute intervals for GLP-1.

Mean glucose levels were significantly lower in both groups during the GLP-1 infusion compared to saline, with an average drop of 2.4 ± 0.6 mM in the IGT subjects ($P<0.02$) and 5.2 ± 0.5 mM in the diabetics ($P<0.0005$). Despite this significant reduction in plasma glucose concentration, mean ISRs were not significantly different during the GLP-1 infusion compared to the saline infusion in both groups (Table 4).

TABLE 4

Mean glucose and ISR responses to

12 hour saline or GLP-1 infusion

ID	Mean glucose 12 hour saline	Mean glucose 12 hour GLP-1	Mean ISR 12 hour saline	Mean ISR 12 hour GLP-1
IGT				
D01	7.49	6.48	376.1	390.9
D02	8.34	6.06	457.9	465.9
D03	10.16	6.46	900.2	1042.7
D04	7.90	6.36	328.5	365.9
D05	10.06	6.37	798.0	1005.8
Mean±SEM	8.79±0.56	6.35±0.08*	572.1±116.1	654.2±152.1
NIDDM				
D06	16.73	11.53	232.3	477.7
D07	17.28	10.51	640.3	715.4
D08	9.94	6.24	615.6	487.7
D09	13.95	8.84	366.5	412.5
D10	15.05	9.90	346.2	448.9
Mean±SEM	14.59±1.3	9.40±0.9*	440.2±80.1	508±53.4

*P<0.05, by paired t test, refers to comparison between saline and GLP-1 infusion.

Mean insulin levels were also maintained during the GLP-1 infusion compared to saline, increasing by 102 ± 90 pmol/L; P=0.32 in subjects with IGT and increasing 7 ± 12 pmol/L; P=0.56 in subjects with NIDDM. Mean glucagon levels were also not different during GLP-1 infusion compared to saline (39.3 ± 5.4 pg/ml vs. 39.4 ± 5.9 pg/ml; P=0.94) in subjects with IGT and (46.4 ± 3.2 pg/ml vs. 42.8 ± 5.4 pg/ml; P=0.4) in subjects with NIDDM. GLP-1 levels achieved during the GLP-1 infusion were 15.6 ± 4.6 pmol/L compared to 2.0 ± 0.8 pmol/L during saline infusion (P<0.001). These levels correspond to postprandial physiological levels.

EXAMPLE 4 (Relationship between glucose and ISR in individual subjects with IGT)

In normal subjects each pulse of glucose is tightly coupled to a pulse in ISR. This coupling has previously been shown to be defective in subjects with IGT. Profiles of glucose and ISR during the oscillatory glucose infusion with saline from one representative subject with IGT, D02, are shown in Figures 3A and 3C. These results demonstrate that in subjects with IGT, during saline infusion, there is loss of the tight coupling between glucose and ISR with many glucose independent oscillations in ISR. In the presence of physiological postprandial levels of GLP-1 (Figures 3B and 3D), the pattern of insulin secretory responses to glucose is improved in the subject with IGT, with each pulse in glucose followed by a pulse in ISR. Hence, GLP-1 improves the ability of the β-cell to entrain an exogenous glucose infusion in the subject with IGT.

EXAMPLE 5 (Relationship between glucose and ISR in individual subjects with NIDDM)

Figure 4 shows the profiles of glucose and ISR from one subject with NIDDM, D07. In marked contrast to subjects with IGT, despite the lowering

of plasma glucose concentrations and the maintenance of ISR, the pattern of insulin secretory responses to glucose was not improved during the GLP-1 infusion (Figures 4B and 4D), with many glucose independent oscillations in ISR persisting. Profiles of glucose and ISR during the oscillatory glucose infusion with saline are shown in Figures 4A and 4C.

EXAMPLE 6 (Effect of GLP-1 on spectral power in IGT and NIDDM)

To determine whether insulin secretion was entrained by glucose in individual subjects, the temporal profiles of insulin secretion were analyzed by spectral analysis. Spectral power analysis was used to evaluate the presence of tight coupling between oscillations in glucose and oscillations in ISR. This method evaluated the regularity of insulin secretory oscillations at a predetermined frequency. Spectral peaks correspond to the dominant periodicity and height of the peaks correspond to spectral power. Each spectrum was normalized assuming the total variance of each series to be 100% and was expressed as the normalized spectral power.

The mean normalized spectral power for glucose in subjects with IGT was 11.2 ± 1.5 during saline infusion and 13.2 ± 1.6 during GLP-1 infusion (P=0.19), and in subjects with NIDDM 6.5 ± 1.8 during saline infusion and 9.8 ± 0.7 during GLP-1 infusion (P=0.18). Figure 5 clearly demonstrates that infusion of GLP-1 in subjects with IGT enhanced insulin secretory responses to the oscillations in plasma glucose, resulting in a greater degree of entrainment of insulin secretion to glucose. The effect was quantified by comparing the normalized spectral power of the insulin secretory profiles. Spectral power for ISR increased from 2.9 ± 1.4 during saline infusion to 8.9 ± 1.7 during GLP-1 infusion; ($P<0.006$) and was unchanged in subjects with NIDDM (1.1 ± 0.5 to 1.5 ± 0.8; P=0.6).

Spectral analysis of the oscillatory glucose profiles confirmed the existence of peaks in the plasma glucose spectra at 144 minutes corresponding to the period of exogenous glucose infusion. Individual power spectra for

glucose and ISR in one subject with IGT (Figure 6A) and one subject with NIDDM (Figure 6B) during saline infusion and during GLP-1 infusion are shown in Figure 6. These correspond to the data shown in Figures 3C and 3D, and Figures 4A and 4B. Spectral power increased from 0.6 to 8.9 in the IGT subject and was minimally changed from 0.28 to 1.51 in the subject with NIDDM. Peaks in plasma glucose spectra occurred at 144 minutes. During saline infusion the dominant spectral peak for ISR did not occur at 144 minutes, but rather was at 0.2. Spectral power with GLP-1 infusion was 1.5.

During saline infusion there was poor entrainment (Fig. 3A) as the spectral power for ISR at 144 was 0.6. During GLP-1 infusion (Fig. 3B) the periodicity of the dominant spectral peak in ISR occurred at 144 minutes, demonstrating that GLP-1 caused the entrainment of the β-cell in this subject.

The mean value for normalized spectral power in historical weight matched control subjects (BMI 28.3) with normal glucose tolerance was 7.2 ± 0.6(9).

The results of this study demonstrated that continuous infusion of physiological postprandial levels of GLP-1 reduced plasma glucose concentrations and stimulated insulin secretion in IGT and NIDDM subjects. Most importantly, GLP-1 restored the ability of the β-cell to sense and respond to plasma glucose in all IGT subjects (quantified by normalized spectral power), with a variable response in subjects who had already developed NIDDM.

The possible mechanisms via which β-cell function is improved by GLP-1 include upregulation of glucose-sensing elements, elimination of glucotoxicity, and improvement in insulin resistance. GLP-1 and glucose exert synergistic insulinotropic actions on β-cells, including stimulation of cyclic AMP formation, insulin secretion, insulin biosynthesis, and proinsulin gene expression.

These examples demonstrated that continuous infusion of physiological levels of GLP-1 reduced plasma glucose concentrations and stimulated insulin secretion in IGT and NIDDM subjects. Most importantly, GLP-1 restored the

ability of the β-cell to sense and respond to small changes in plasma glucose concentrations in IGT subjects, with only a variable response in NIDDM subjects. In IGT subjects, we observed a significant increase in spectral power, a measure of β-cell function that does not rely on adjustment for changes in insulin sensitivity.

What is claimed is:

1. A composition comprising a compound which binds to a receptor for "GLP-1", and a pharmaceutical carrier, said compound being present in an amount effective to enhance the sensitivity and response of pancreatic β-cells to changes in plasma glucose, as measured by the timing and amount of insulin secretions in response to increases in plasma glucose, in a human with impaired glucose tolerance.

2. The composition of claim 1 wherein the receptor-binding compound is selected from (a) a peptide which comprises the amino acid sequence of glucagon-like peptide-1, and (b) a variant peptide comprising an amino acid sequence that differs from the sequence of glucagon-like peptide-1 by one or more substitutions, deletions or insertions.

3. The composition of claim 2 wherein the receptor binding compound is glucagon-like peptide-1.

4. The compound of claim 2 wherein the receptor binding compound is glucagon-like peptide-1 (7-37) which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:3).

5. The compound of claim 2 wherein the receptor binding compound is glucagon-like peptide-1 (7-36) amide which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:4).

6. The composition of claim 2 wherein the receptor binding compound is a variant peptide in which the combination of the substitutions, deletions and

insertions in the amino acid sequence does not differ by more than ten amino acids from the amino acid sequence of glucagon-like peptide-1.

7. The composition of claim 1, further comprising an agent which enhances the half-life *in vivo* of the compound.

8. The composition of claim 1 wherein the receptor binding compound is expressed by a polynucleotide.

9. The composition of claim 1 wherein the receptor binding compound is an organic molecule having a molecular weight of not greater than about 5000.

10. A method for treating an individual with impaired glucose tolerance comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to enhance the regularity of insulin responses, and the amplitude thereof, in reaction to changes in plasma glucose.

11. The method of claim 10 wherein the receptor binding compound is selected from (a) a peptide which comprises the amino acid sequence of glucagon-like peptide-1, and (b) a variant peptide comprising an amino acid sequence that differs from the sequence of glucagon-like peptide-1 by one or more substitutions, deletions or insertions.

12. The method of claim 11 wherein the receptor binding compound is glucagon-like peptide-1.

13. The method of claim 11 wherein the receptor binding compound is glucagon-like peptide-1 (7-37) which has the sequence His Ala Glu Gly Thr

Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:3).

14. The method of claim 11 wherein the receptor binding compound is glucagon-like peptide-1 (7-36) amide which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:4).

15. The method of claim 11 wherein the receptor binding compound is a variant peptide in which the combination of the substitutions, deletions and insertions in the amino acid sequence does not differ by more than ten amino acids from the amino acid sequence of glucagon-like peptide-1.

16. The method of claim 10 wherein the receptor binding compound is expressed by a polynucleotide.

17. The method of claim 10 wherein the receptor binding compound is an organic molecule having a molecular weight of not greater than about 5000.

18. The method of claim 10 wherein the step of administering is selected from the group consisting of intravenous, subcutaneous, intramuscular, interperiotoneal, injected depot with sustained release, deep lung insufflation with sustained release, buccal or patch.

19. The method of claim 10, further comprising administering an agent that enhances the half-life *in vivo* of said receptor binding compound.

20. The method of claim 19 wherein the agent is administered concurrently with the composition.

21. The method of claim 19 wherein the agent is covalently linked to the receptor binding compound.

22. The method of claim 18 wherein intravenous administration is in a dose range of from about 0.3 to about 2.0 pmol/kg per minute.

23. The method of claim 18 wherein continuous subcutaneous administration is in a dose range of from about 1.0 to about 20.0 pmol/kg per minute.

24. A method for treating a human with impaired glucose tolerance, comprising: administering to the human a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to retard or arrest the loss of plasma glucose control and the development of non-insulin dependent diabetes mellitus.

25. The method of claim 24 wherein the receptor binding compound is selected from (a) a peptide which comprises the amino acid sequence of glucagon-like peptide-1, and (b) a variant peptide comprising an amino acid sequence that differs from the sequence of glucagon-like peptide-1 by one or more substitutions, deletions or insertions.

26. The method of claim 25 wherein the receptor binding compound is glucagon-like peptide-1.

27. The method of claim 25 wherein the receptor binding compound is glucagon-like peptide-1 (7-37) which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg Gly (SEQ. ID NO:3).

28. The method of claim 25 wherein the receptor binding compound is glucagon-like peptide-1 (7-36) amide which has the sequence His Ala Glu Gly Thr Phe Thr Ser Asp Val Ser Ser Tyr Leu Glu Gly Gln Ala Ala Lys Glu Phe Ile Ala Trp Leu Val Lys Gly Arg (NH_2) (SEQ. ID NO:4).

29. The method of claim 25 wherein the receptor binding compound is a variant peptide in which the combination of the substitutions, deletions and insertions in the amino acid sequence does not differ by more than five amino acids from the amino acid sequence of glucagon-like peptide-1.

30. The method of claim 24 wherein the receptor binding compound is expressed by a polynucleotide.

31. The method of claim 24 wherein the receptor binding compound is an organic molecule having a molecular weight of not greater than about 5000.

32. The method of claim 24 wherein the step of administering is selected from the group consisting of intravenous, subcutaneous, intramuscular, interperitoneal, injected depot with sustained release, deep lung insufflation with sustained release, buccal or patch.

33. The method of claim 32 wherein intravenous administration is in a dose range of from about 0.1 to about 10.0 pmol/kg per minute.

34. The method of claim 32 wherein continuous subcutaneous administration is in a dose range of from about 0.1 to about 75.0 pmol/kg per minute.

35. A method for treating an individual with impaired glucose tolerance comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a

pharmaceutical carrier, wherein said composition contains an amount of said compound effective to improve entrainment of β-cell insulin secretory responses to exogenous glucose oscillations.

36. A method for treating an individual with impaired glucose tolerance comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to enhance a normalization of insulin secretory patterns in impaired glucose tolerance.

37. A method for treating an individual with impaired glucose tolerance comprising: administering to said individual a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to reduce plasma insulin levels in an individual with impaired glucose tolerance.

38. A method for treating an individual with impaired glucose tolerance comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to reduce insulin resistance in an individual with impaired glucose tolerance.

39. A method for treating an individual whose symptoms indicate increased risk of a cardiovascular event comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to enhance the regularity of insulin responses, and the amplitude thereof, in reaction to changes in plasma glucose, and to reduce plasma insulin levels.

40. A method for treating an individual whose symptoms indicate increased risk of a cerebrovascular event comprising: administering to said individual a composition comprising a compound which binds to a receptor for glucagon-like peptide-1 and a pharmaceutical carrier, said composition containing an amount of said compound effective to enhance the regularity of insulin responses, and the amplitude thereof, in reaction to changes in plasma glucose, and to reduce plasma insulin levels.

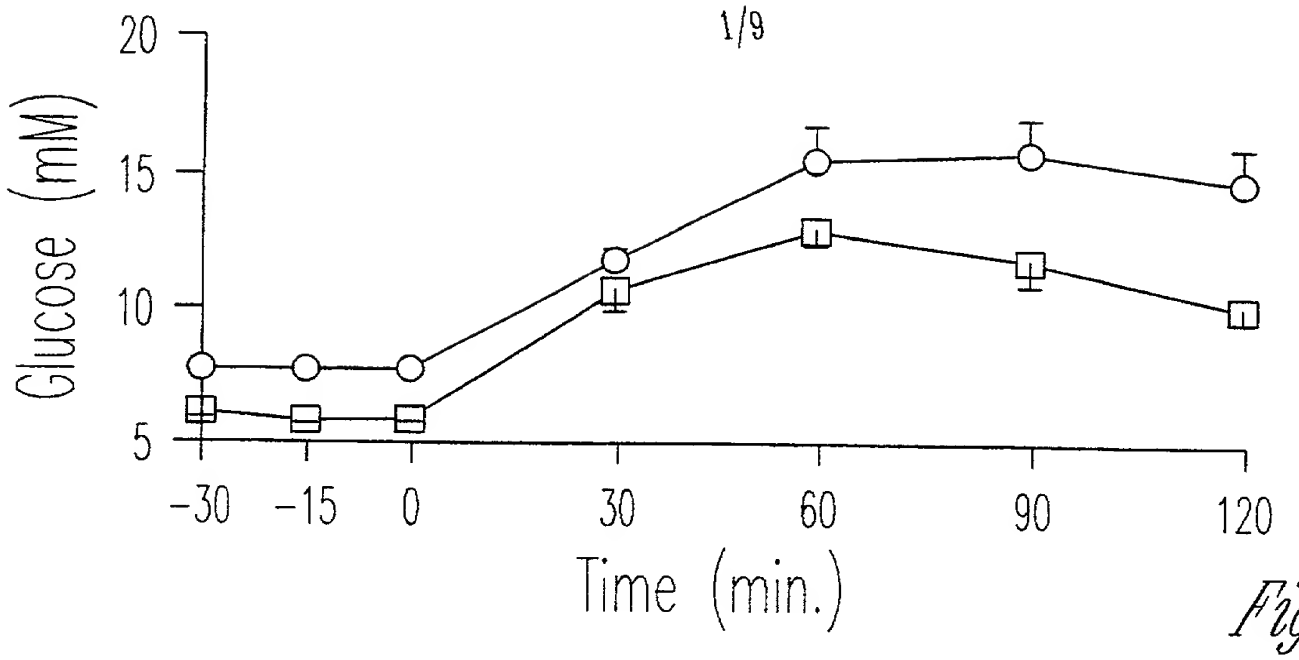


Fig. 1A

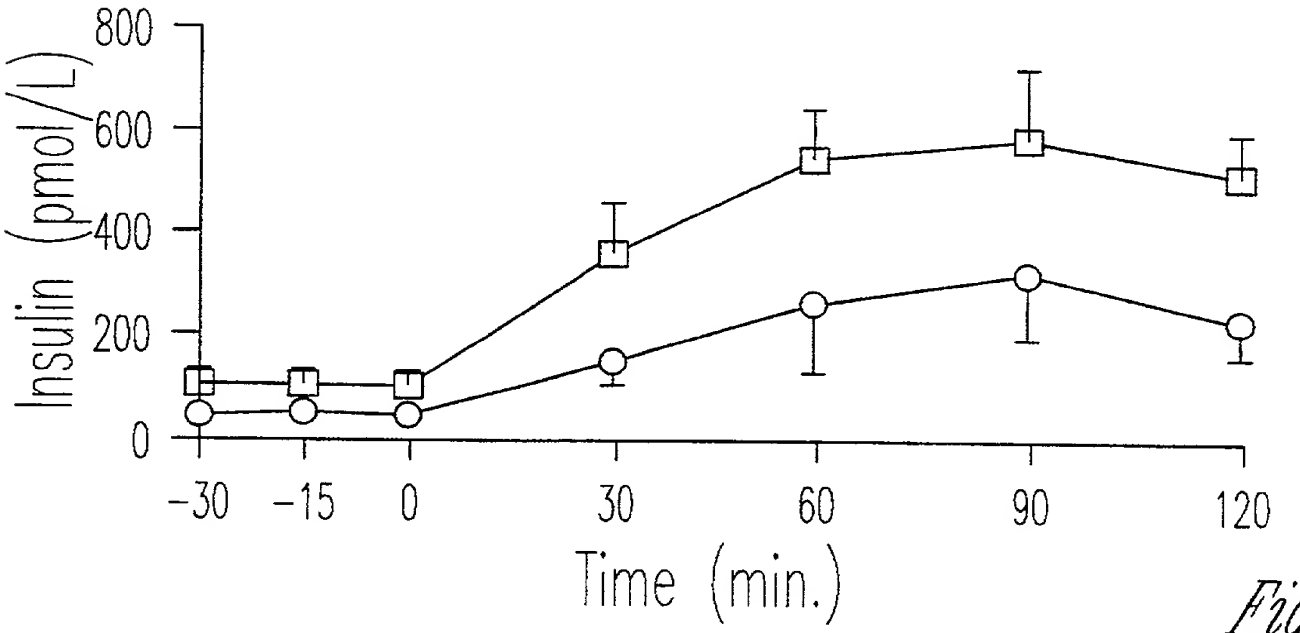


Fig. 1B

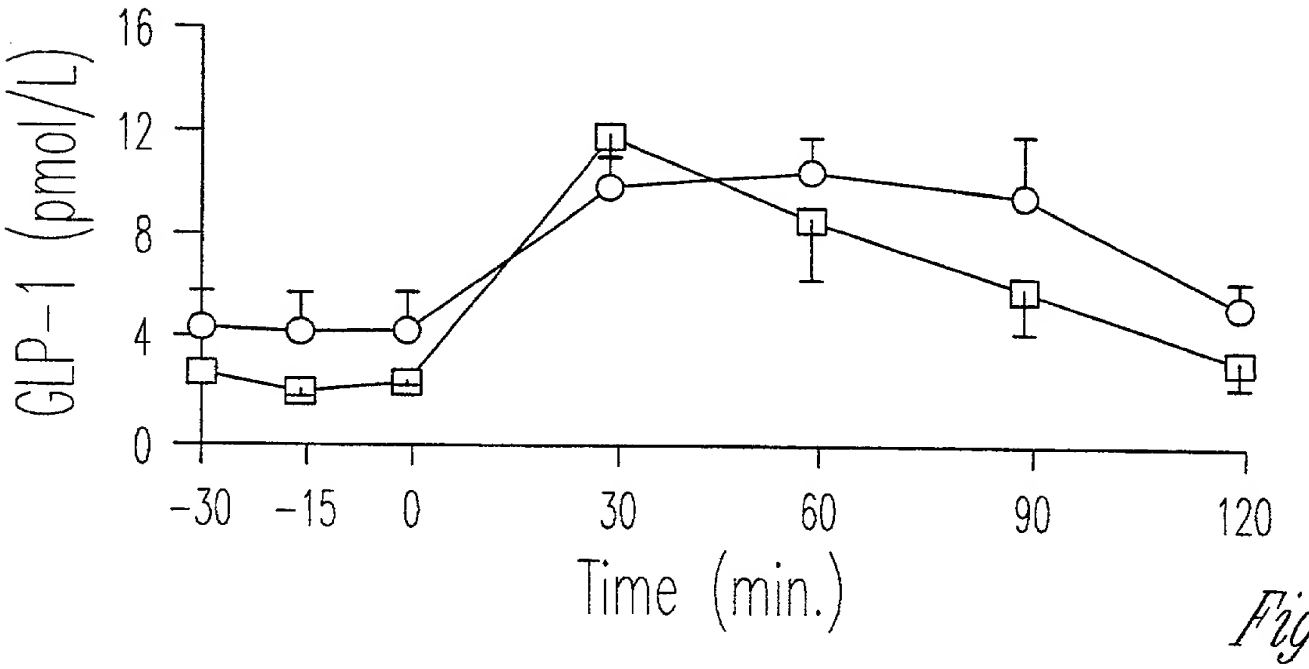
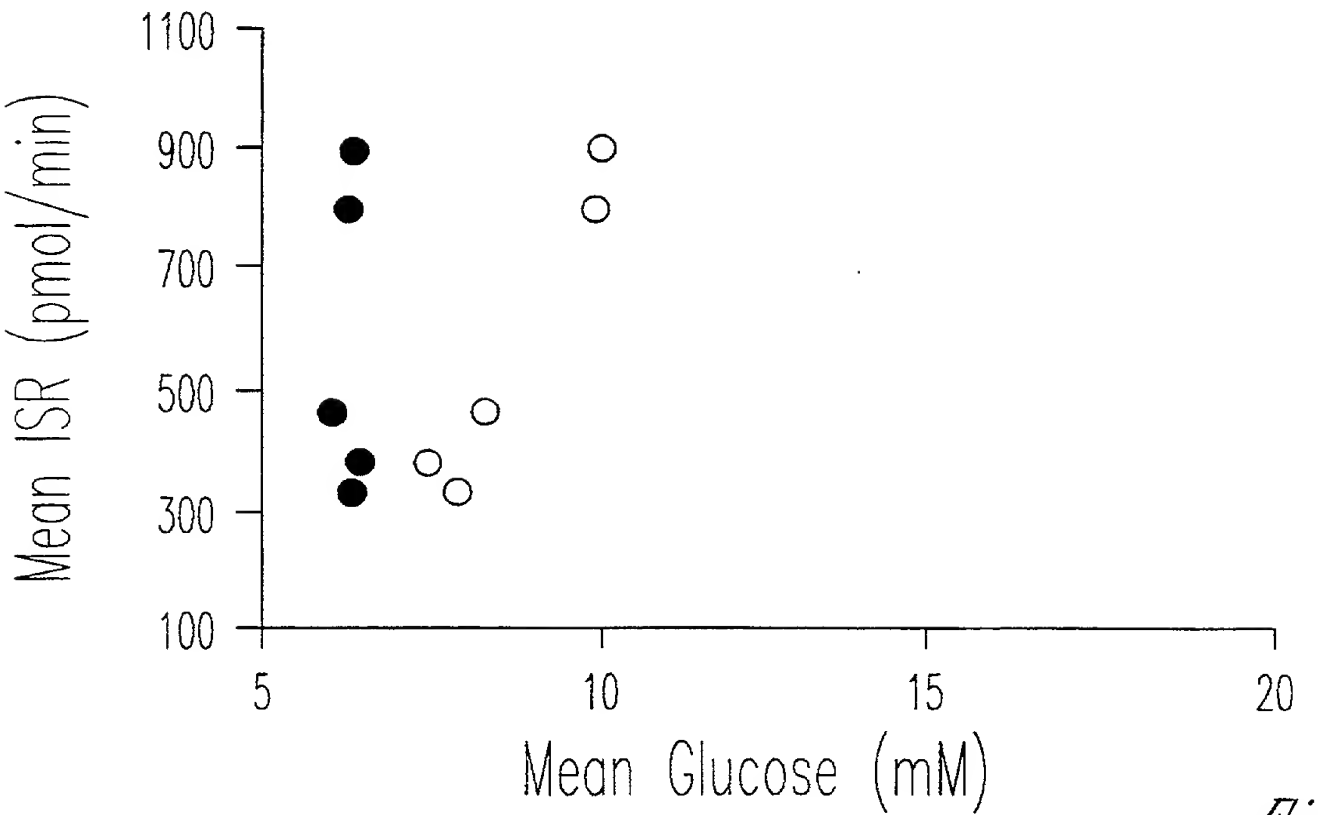


Fig. 1C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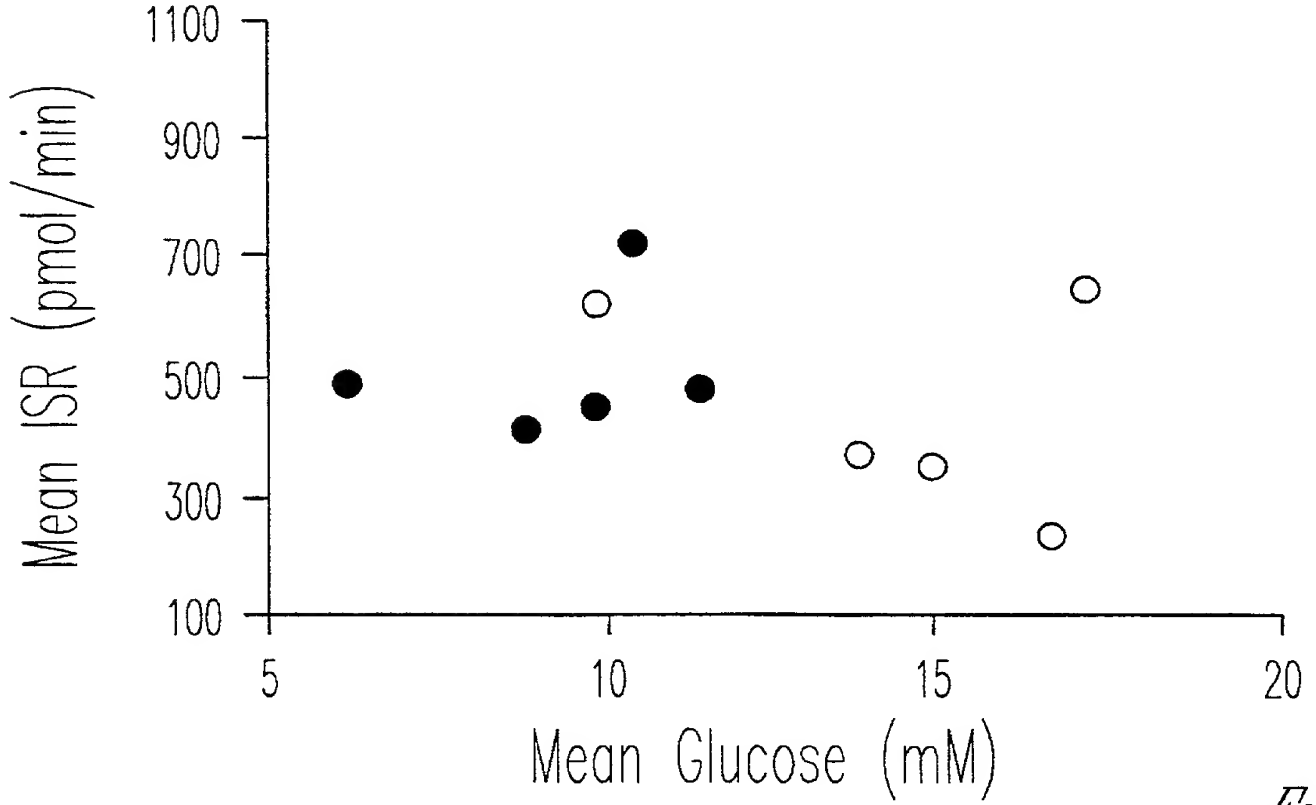
700
500
300
100
Mean ISR (pmol/min)
5
10
15
20
Mean Glucose (mM)

Fig. 2A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300
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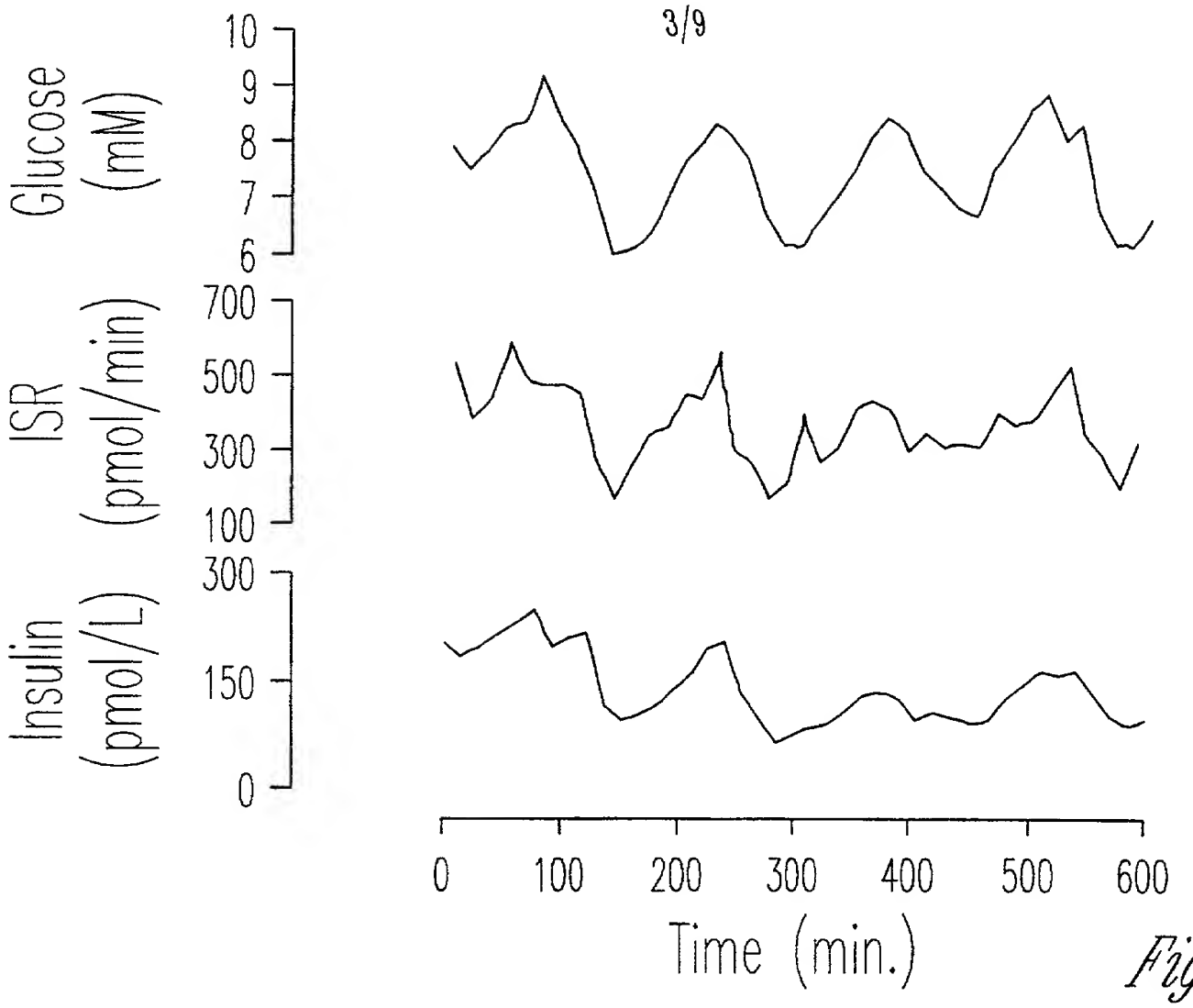
Mean ISR (pmol/min)
5
10
15
20
Mean Glucose (mM)

Fig. 2B


Glucose (mM)
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9
8
7
6
ISR (pmol/min)
700
500
300
100
Insulin (pmol/L)
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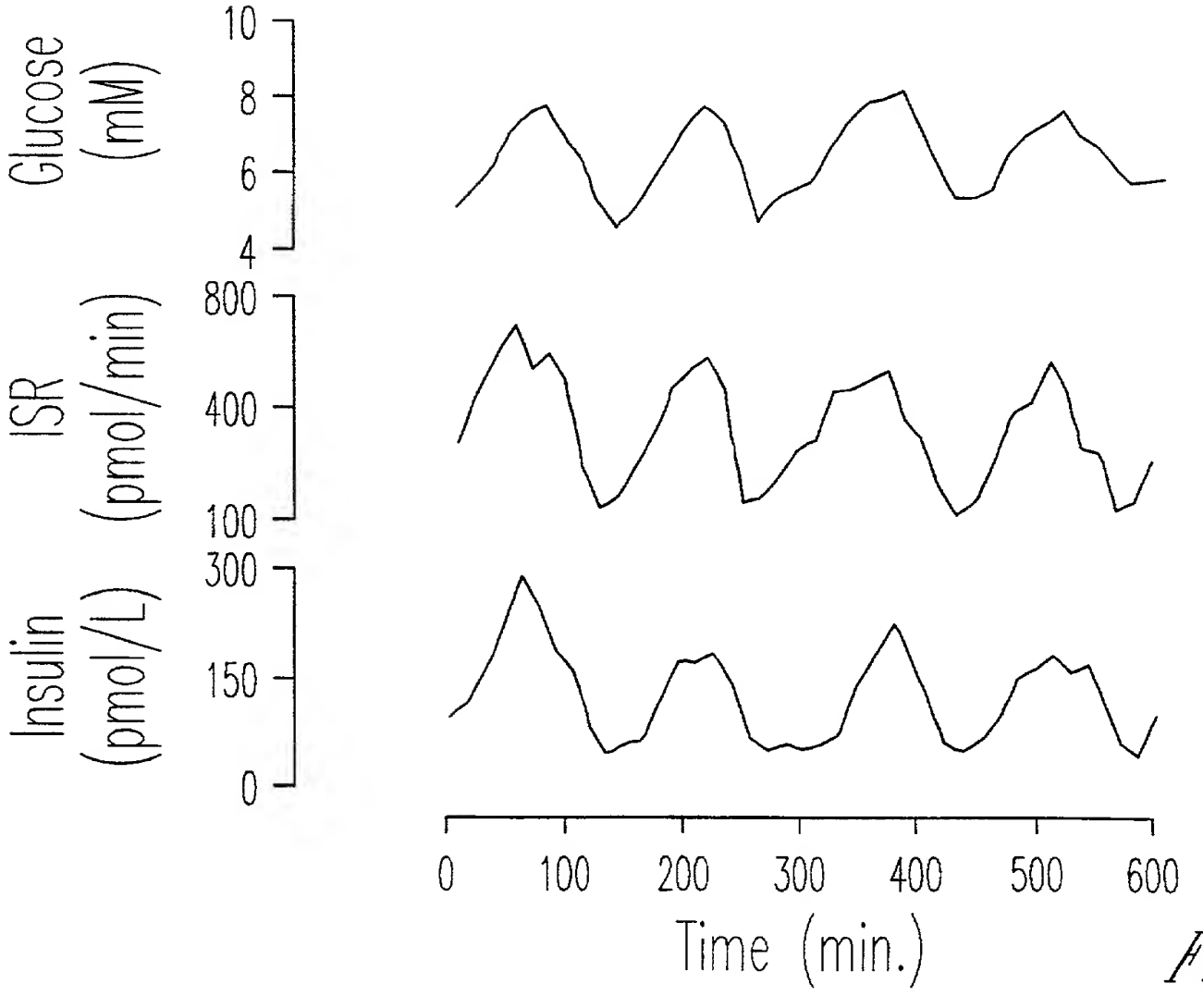
0
100
200
300
400
500
600
Time (min.)

Fig. 3A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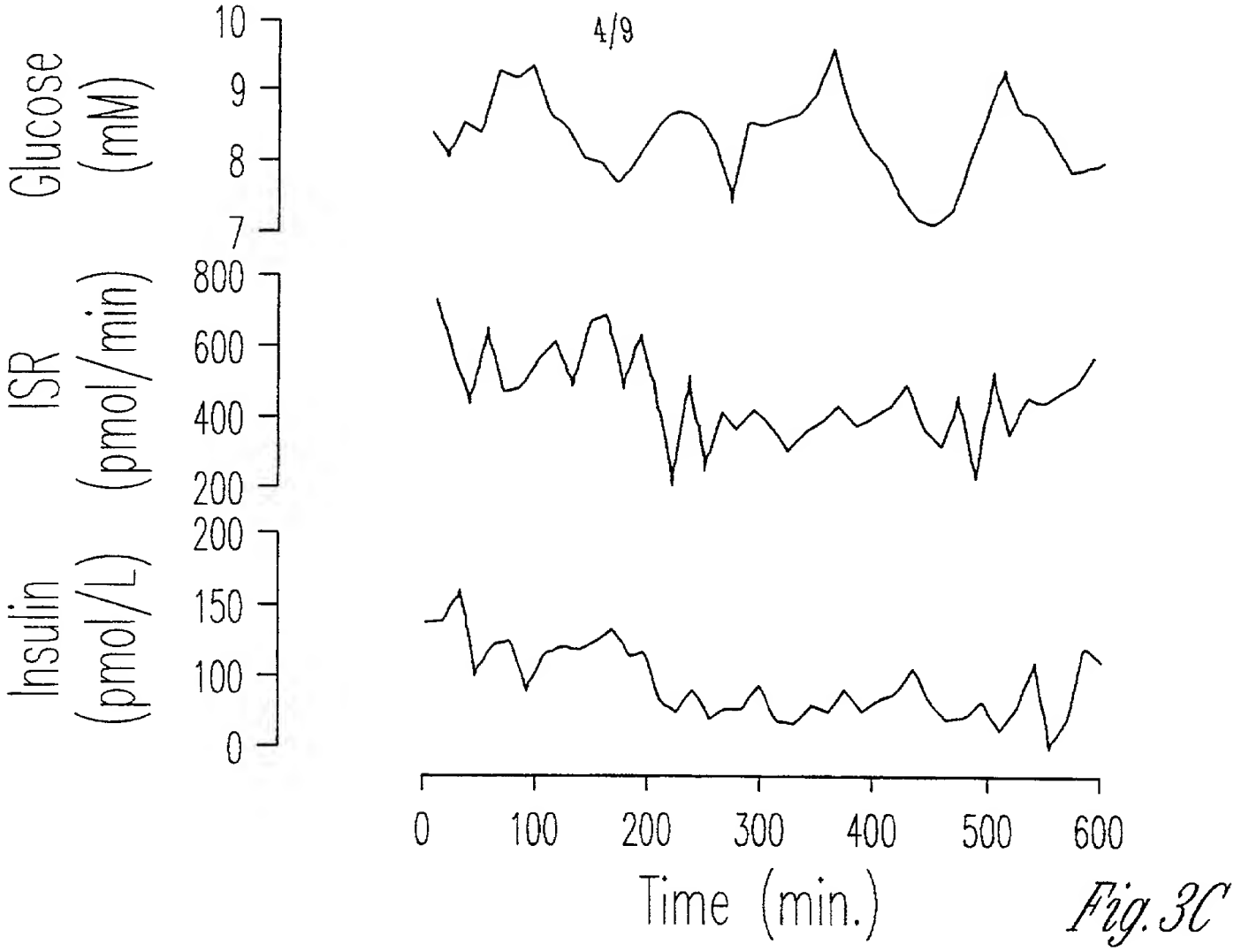
400
500
600
Time (min.)

Fig. 3B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9
8
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ISR (pmol/min)
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Insulin (pmol/L)
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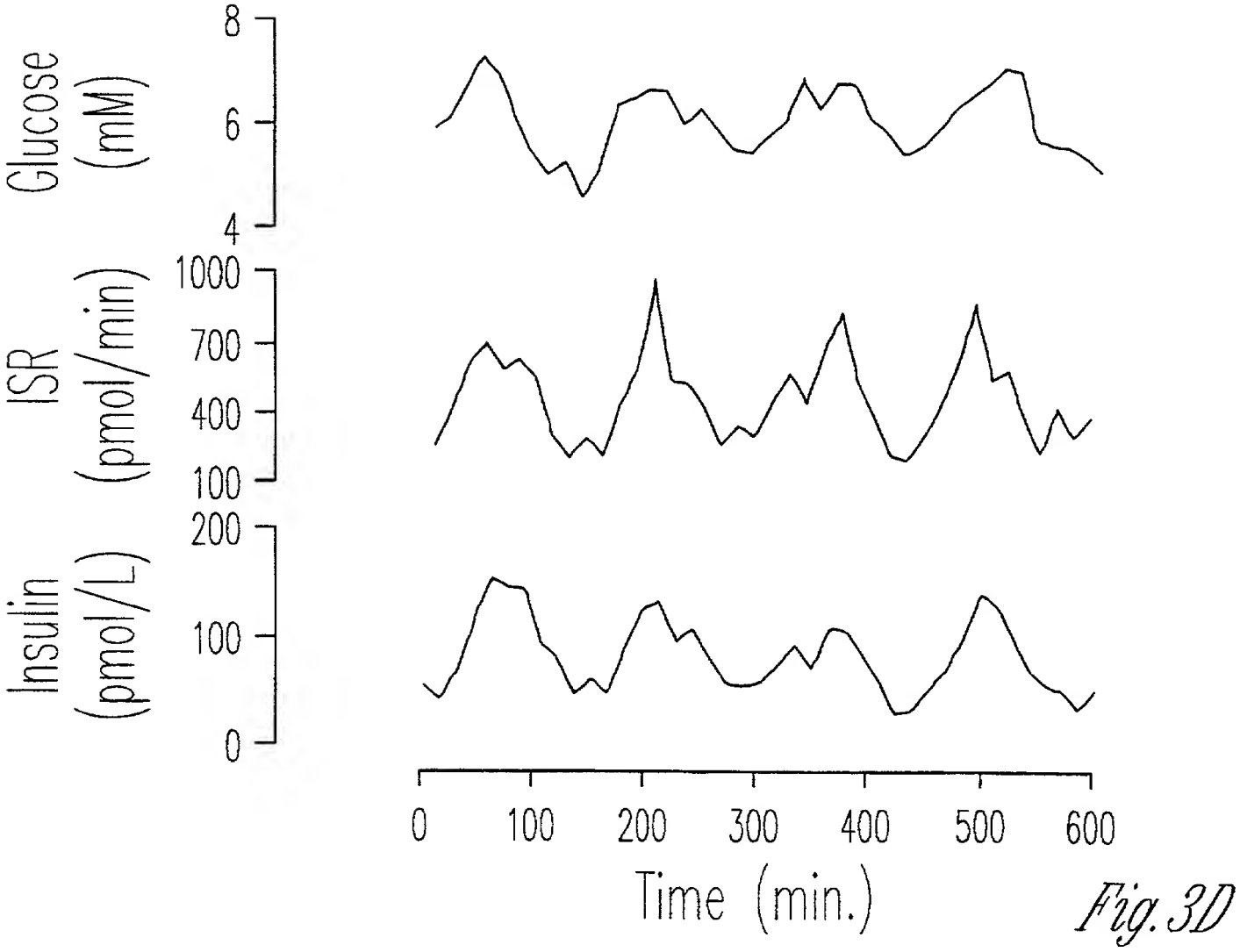
0
100
200
300
400
500
600
Time (min.)

Fig. 3C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4
ISR (pmol/min)
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400
100
Insulin (pmol/L)
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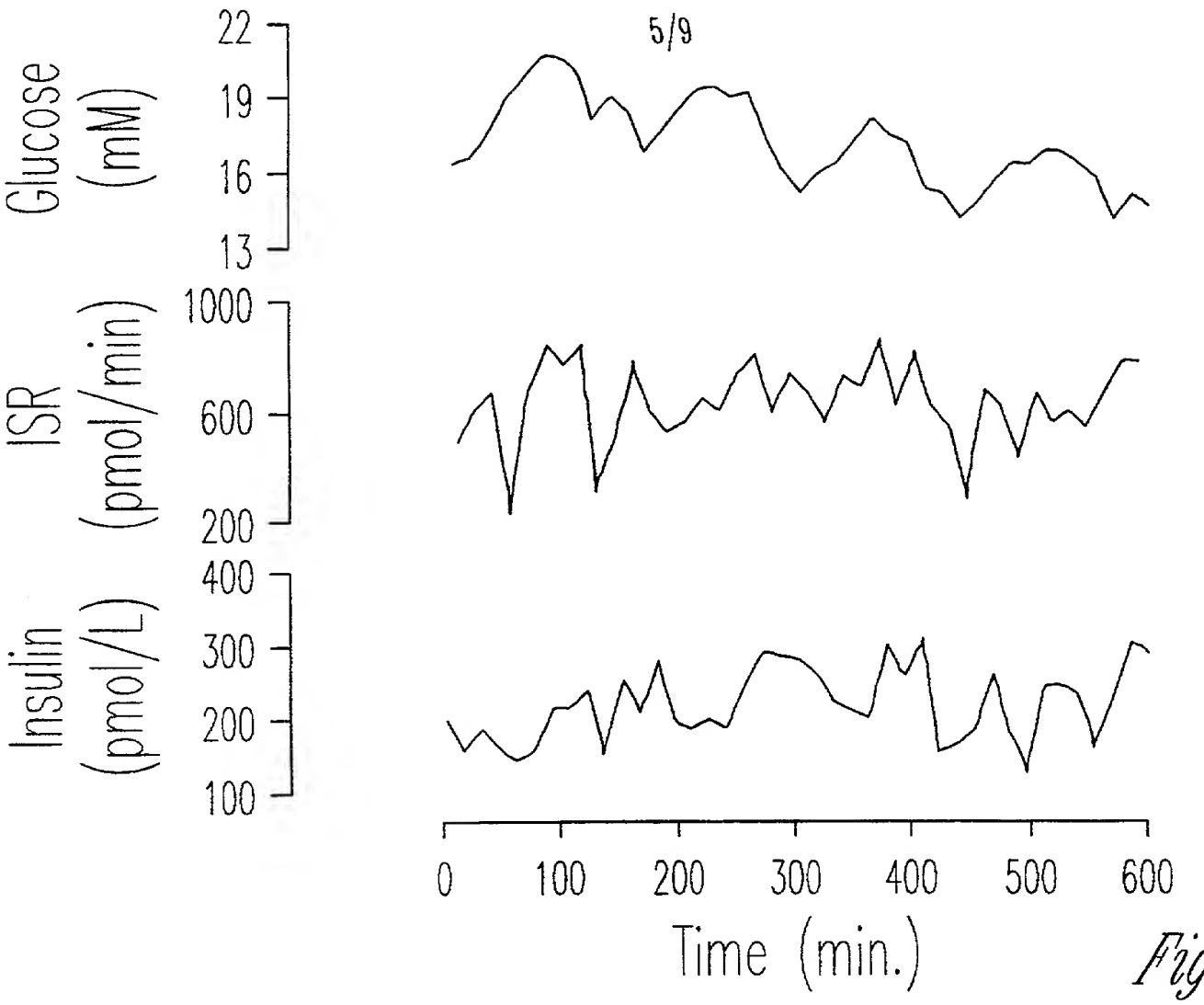
100
0
0
100
200
300
400
500
600
Time (min.)

Fig. 3D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ISR (pmol/min)
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600
200
Insulin (pmol/L)
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200
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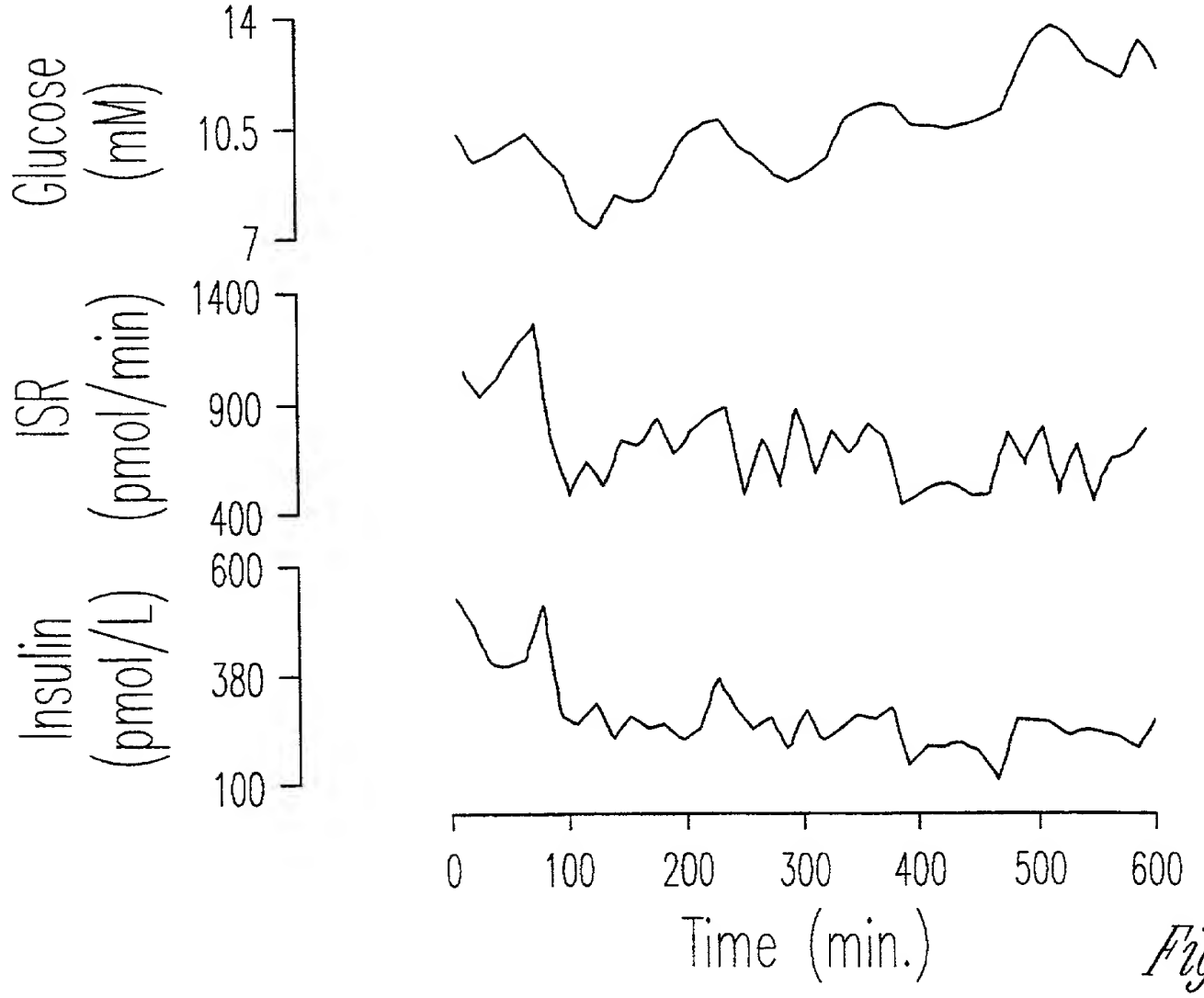
0
100
200
300
400
500
600
Time (min.)

Fig. 4A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ISR (pmol/min)
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900
400
Insulin (pmol/L)
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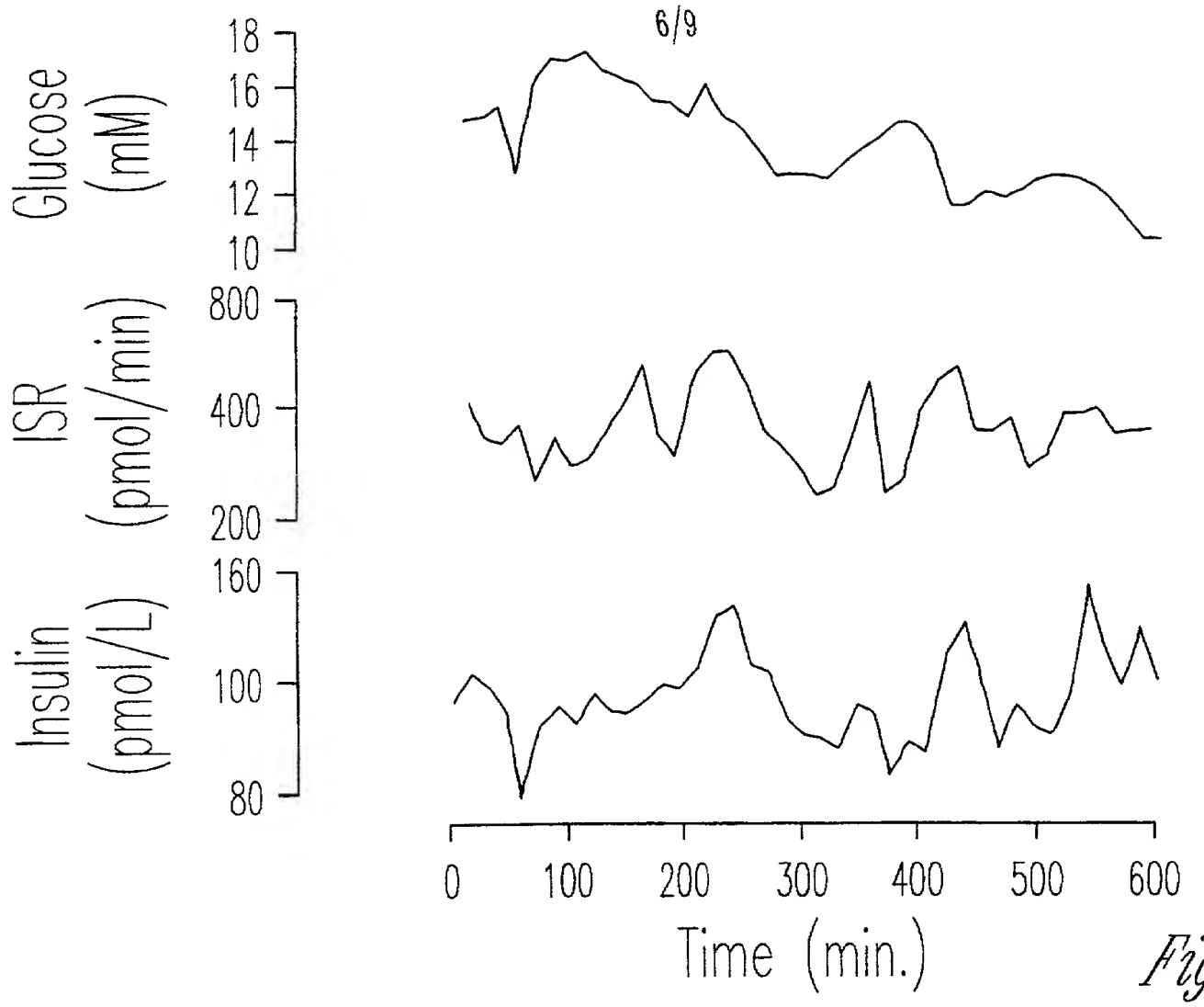
100
0
100
200
300
400
500
600
Time (min.)

Fig. 4B


Glucose (mM)
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16
14
12
10
ISR (pmol/min)
800
400
200
Insulin (pmol/L)
160
100
80
0
100
200
300
400
500
600
Time (min.)

Fig. 4C

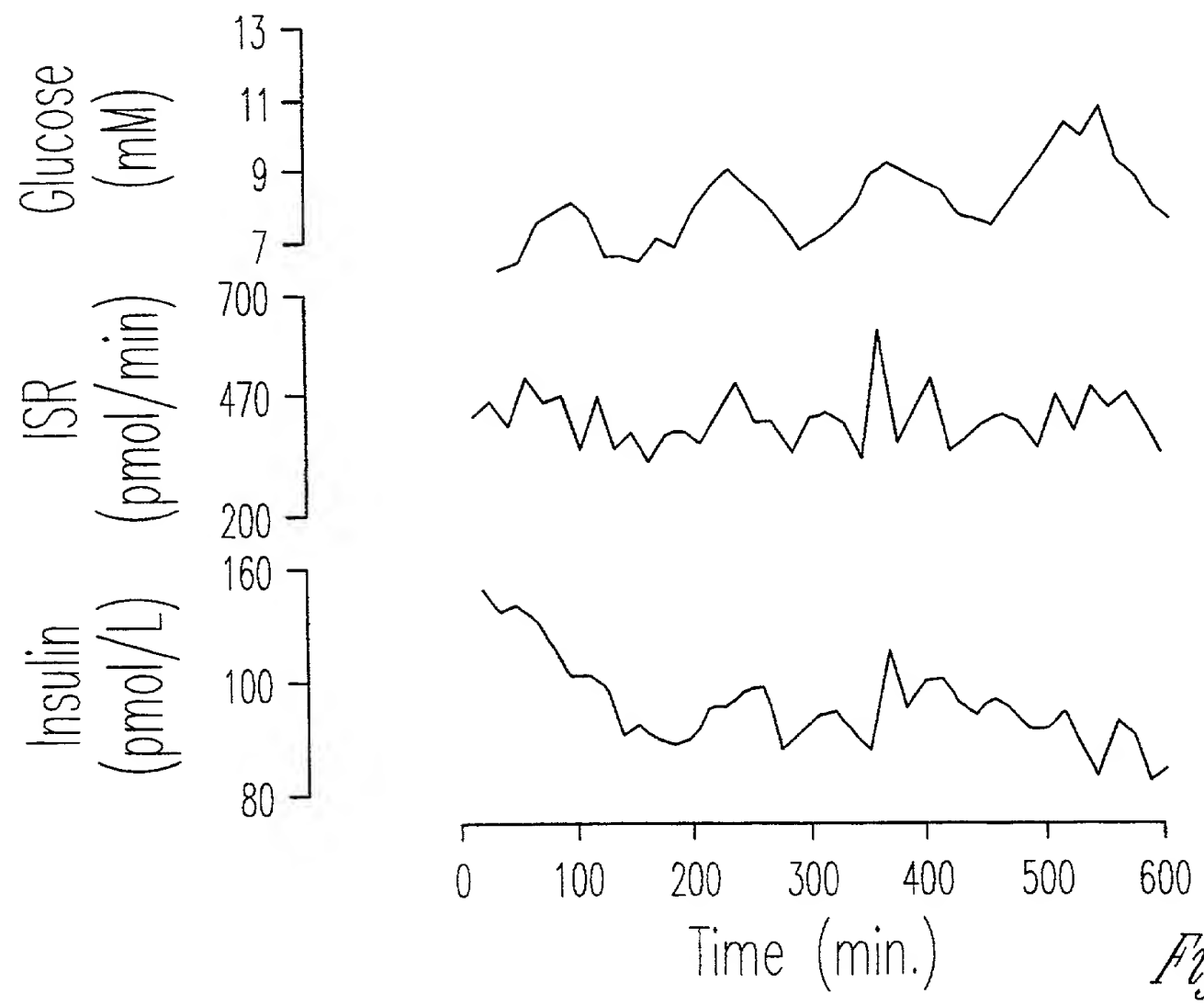
Glucose (mM)
13
11
9
7
ISR (pmol/min)
700
470
200
Insulin (pmol/L)
160
100
80
0
100
200
300
400
500
600
Time (min.)

Fig. 4D

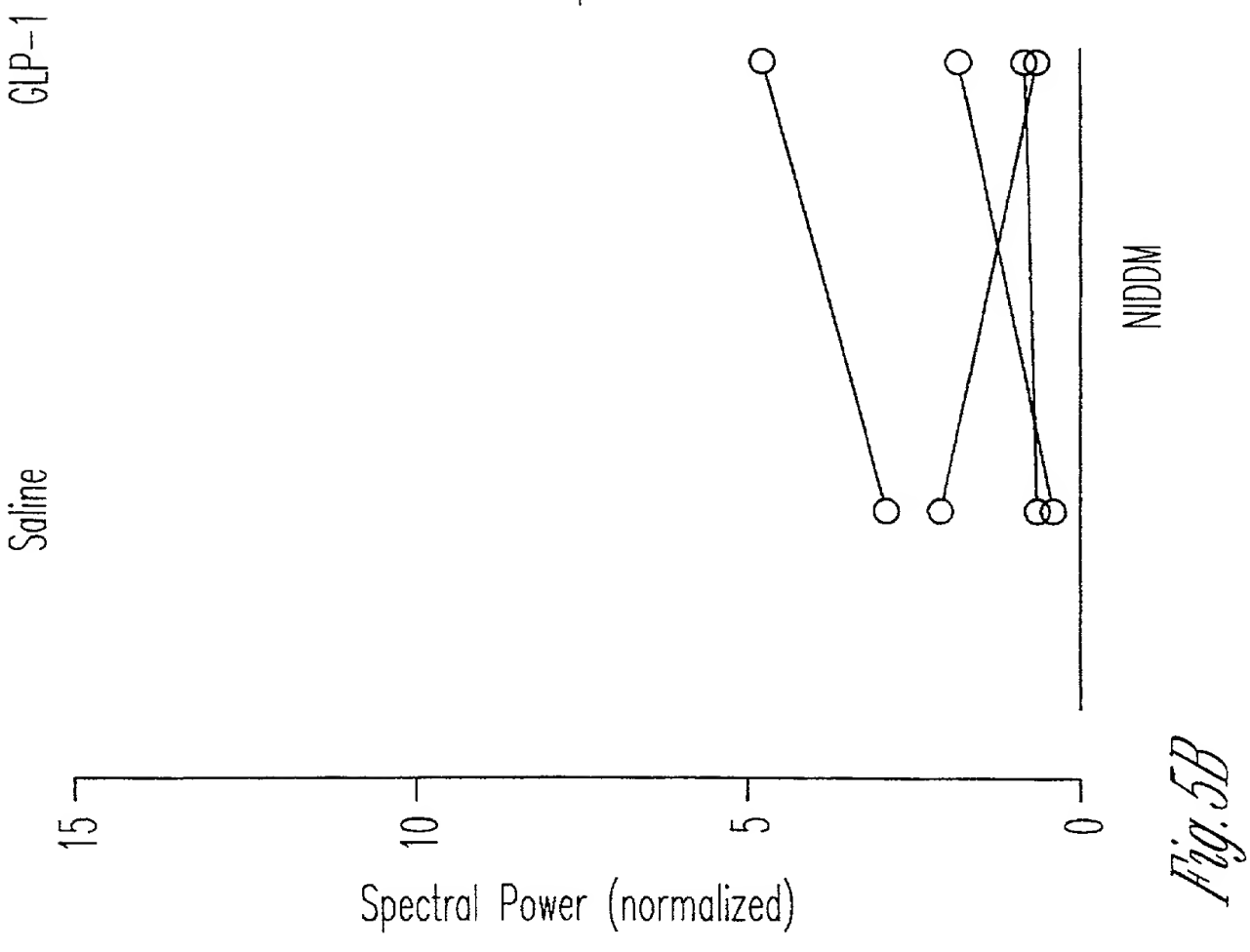
GLP-1
Saline
NIDDM
15
10
5
0
Spectral Power (normalized)

Fig. 5B

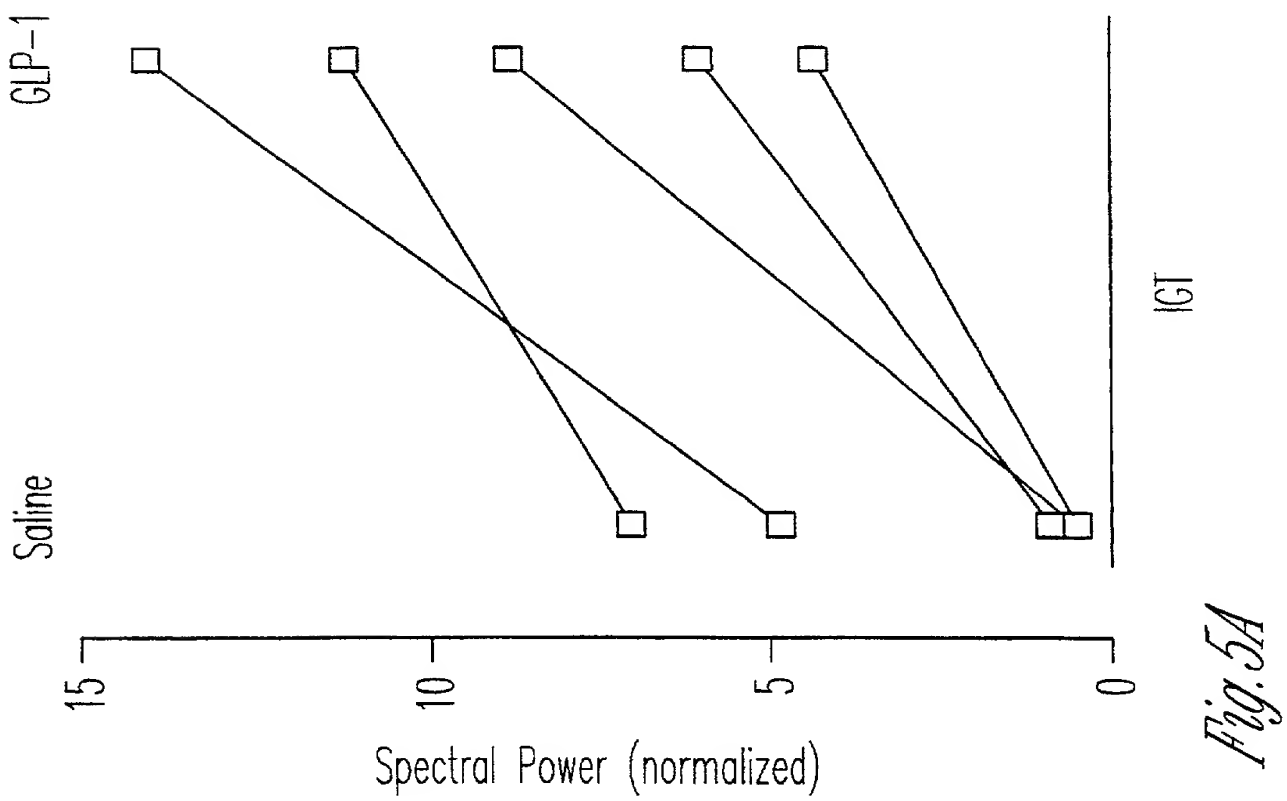
GLP-1
Saline
IGT
15
10
5
0
Spectral Power (normalized)

Fig. 5A

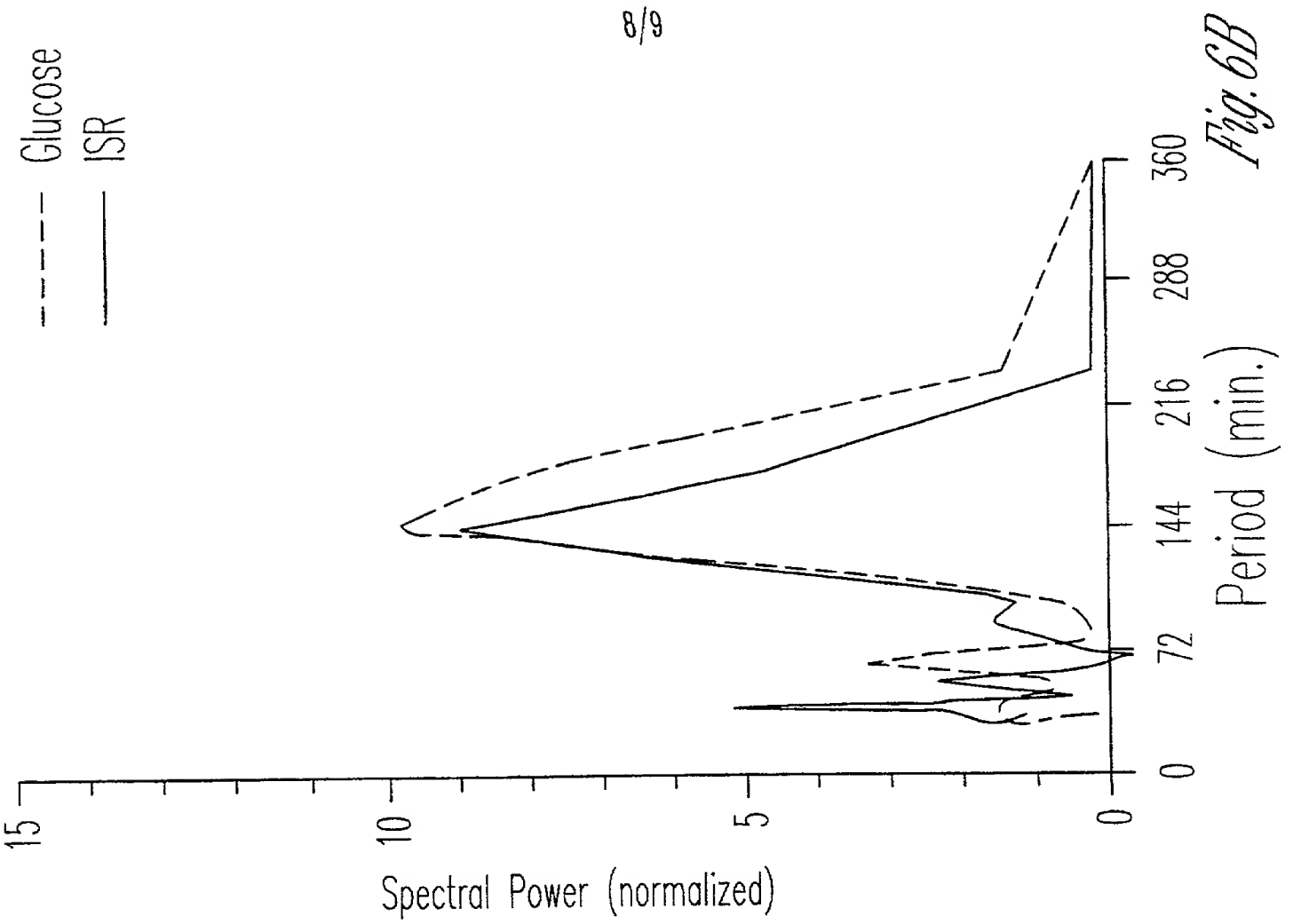
Glucose
ISR
Spectral Power (normalized)
15
10
5
0
0
72
144
216
288
360
Period (min.)

Fig. 6B

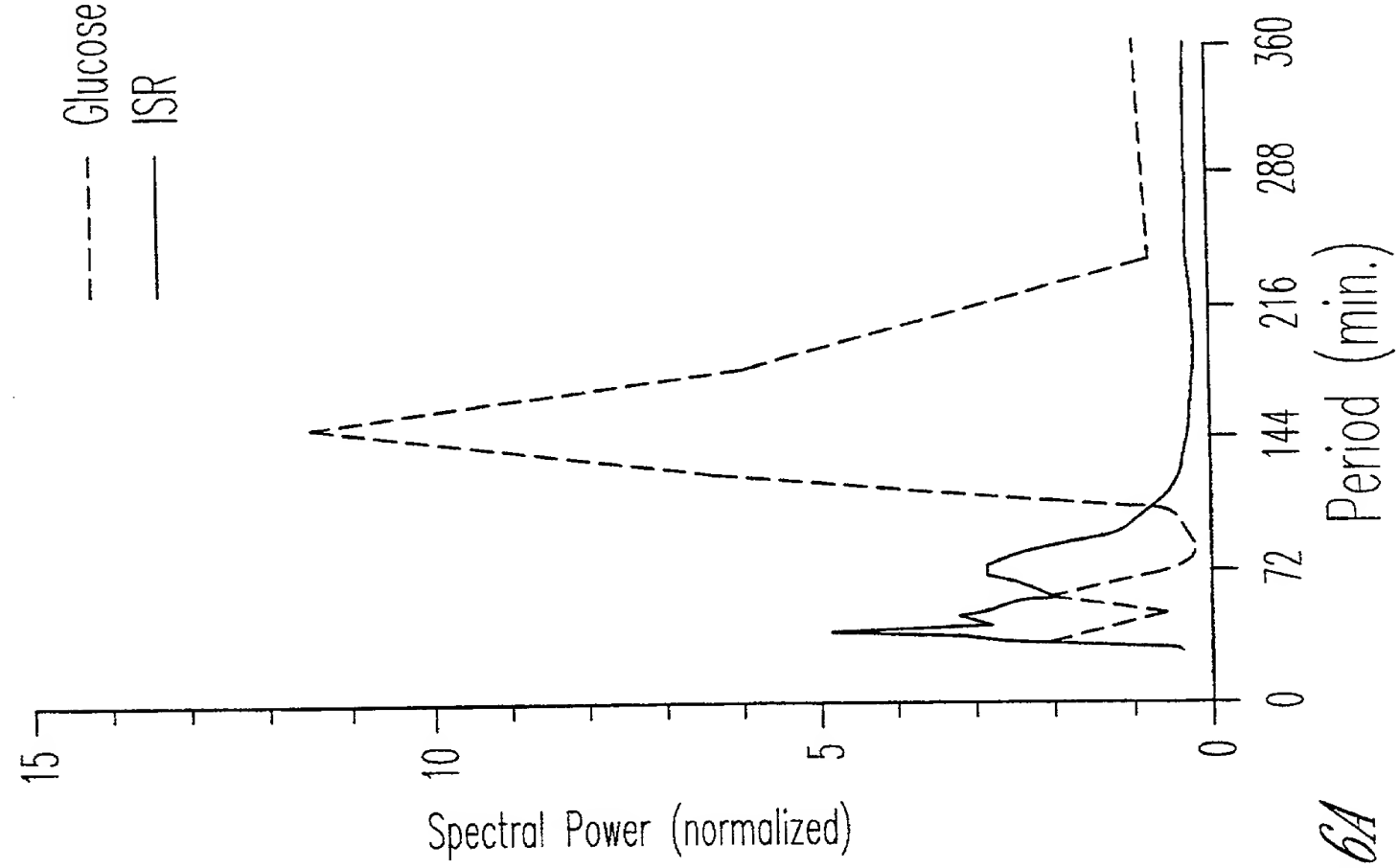
Glucose
ISR
Spectral Power (normalized)
15
10
5
0
0
72
144
216
288
360
Period (min.)

Fig. 6A

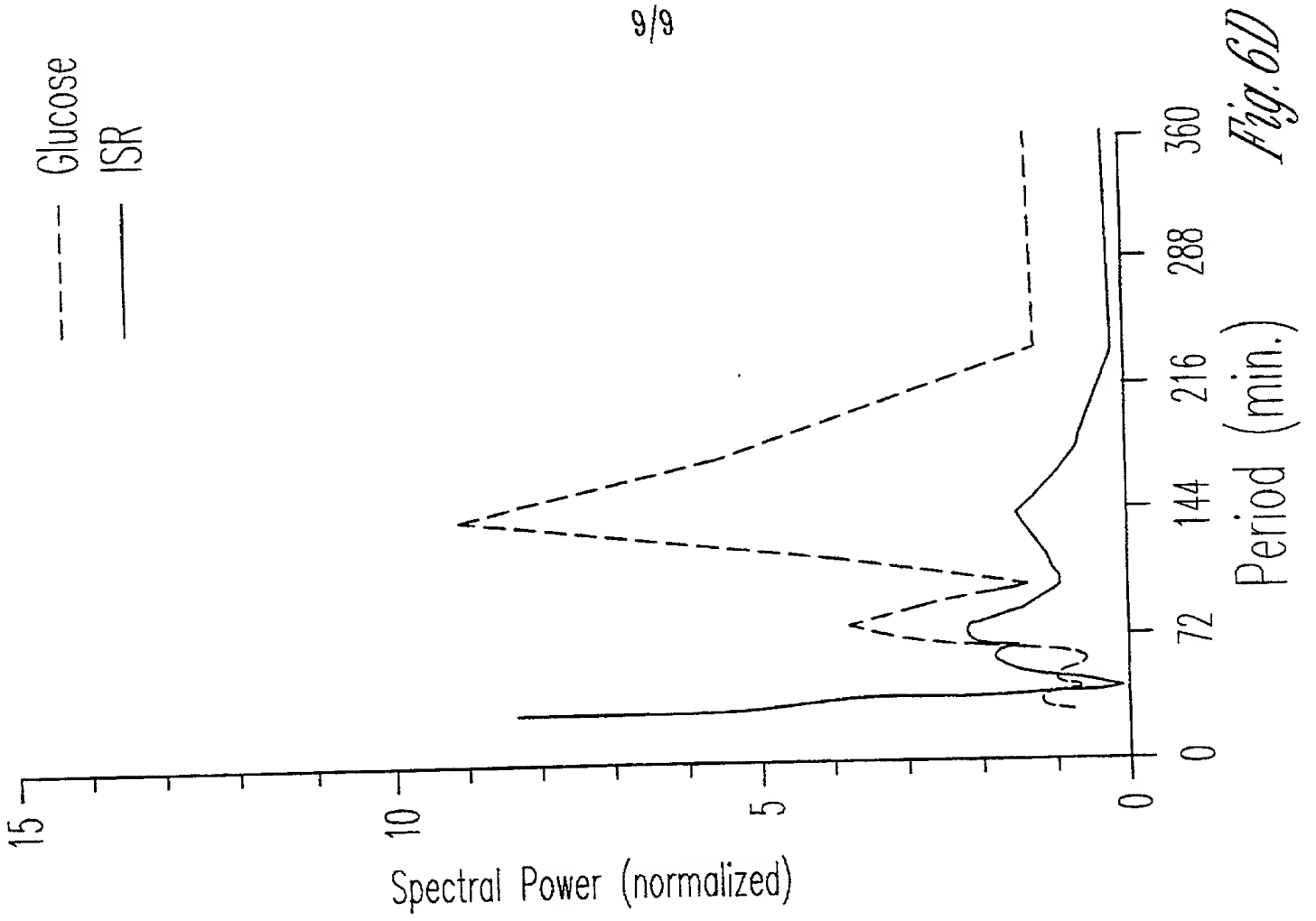
Glucose
ISR
Spectral Power (normalized)
15
10
5
0
0
72
144
216
288
360
Period (min.)

Fig. 6D

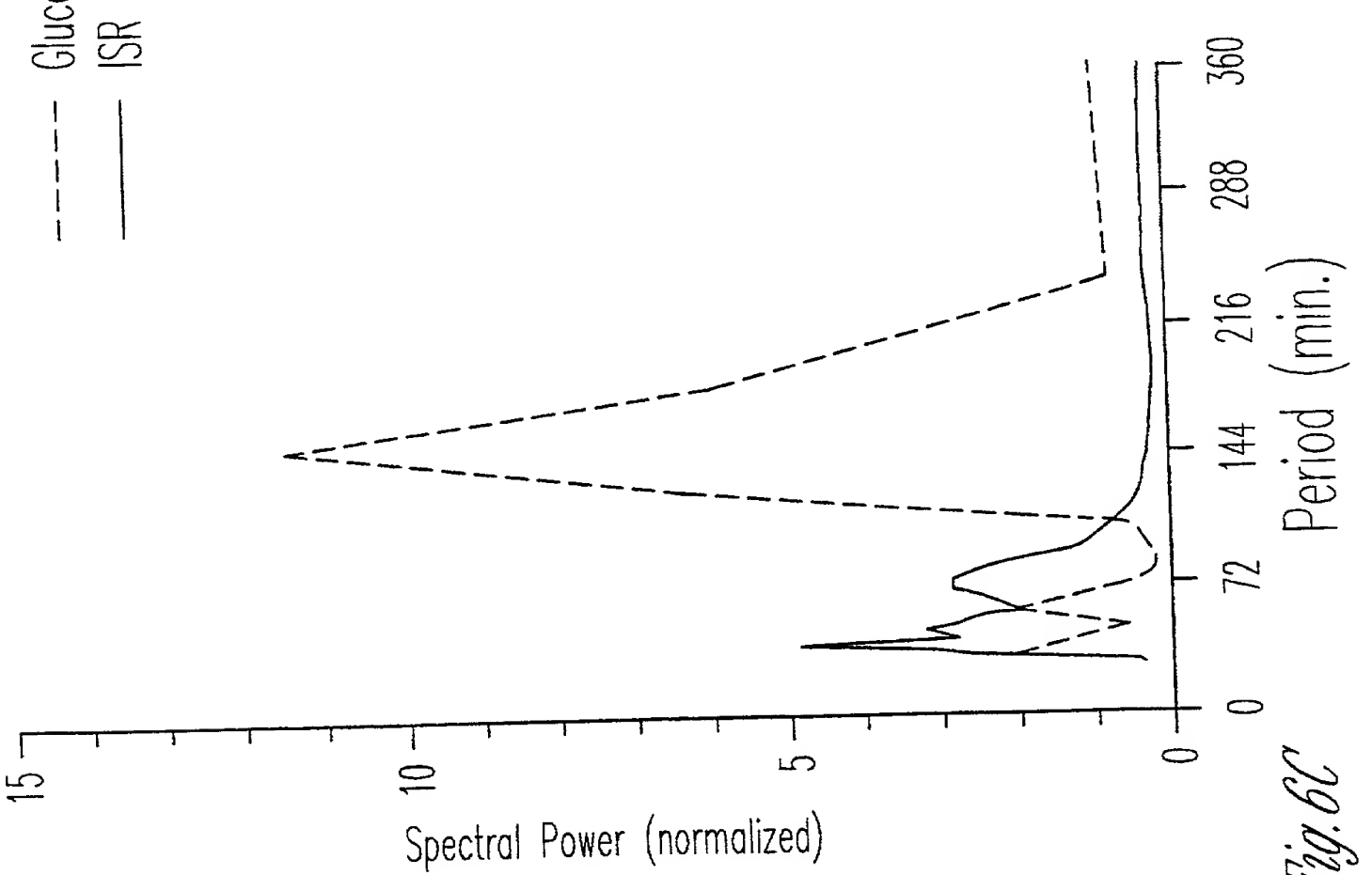
Glucose
ISR
Spectral Power (normalized)
15
10
5
0
0
72
144
216
288
360
Period (min.)

Fig. 6C

Combined Declaration for Patent Application and Power of Attorney (Includes Reference to PCT International Applications)	Attorney's Docket Number P03986US2

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

GLUCAGON-LIKE PEPTIDE-1 IMPROVES THE ABILITY OF THE ß-CELL TO SENSE AND RESPOND TO GLUCOSE IN SUBJECTS WITH IMPAIRED GLUCOSE TOLERANCE

the specification of which (check only one item below)

is attached hereto.

☒ was filed as United States application

Serial No. 09/719,410

on December 12, 2000

and was amended

on _____(if applicable).

☒ was filed as PCT international application (This is the only place we put in info re the parent PCT appl.

Number PCT/US99/10040

on MAY 7, 1999

and was amended under PCT Article 19

on _____(if applicable).

☐

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

☐

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations §1.56(a).

☐

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119: foreign priority

COUNTRY (if PCT indicate PCT)	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 USC 119
			YES NO

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112. I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120: This is for CIP infor, do not put this PCT in here, only prior, if any

U.S. APPLICATIONS			STATUS *(Check one)*		
U.S. APPLICATION NUMBER	U.S. FILING DATE		PATENTED	PENDING	ABANDONED
PCT APPLICATIONS DESIGNATING THE U.S					
PCT APPLICATION NO.	PCT FILING DATE	U.S. SERIAL NUMBERS ASSIGNED *(if any)*			

Applicant or Patentee: Burkhard Goke & Maria Byrne
Serial No. or Patent No: _____
Filed or Issued: _____
For: Glucagon-Like Peptide-1 Improves The Ability Of The B-Cell To Sense And Respond To Glucose In Subjects With Impaired Glucose Tolerance

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS (37 CFR 1.9(f) AND 1.27(c)) - SMALL BUSINESS CONCERN

I hereby declare that I am

[ ] the owner of the small business concern identified below:
[X] an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN BioNebraska, Inc.
ADDRESS OF CONCERN 3820 N.W. 46th Street, Lincoln, NE 68524

I hereby declare that the above-identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention, entitled Glucagon-Like Peptide-1 Improves The Ability of ... by inventor(s) Burkhard Goke; Maria Byrne, described in

[X] the specification filed herewith.
[ ] application Serial No. _____, filed _____.
[ ] Patent No. _____, issued _____.

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights in the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37 CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

*NOTE: *Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27).*

FULL NAME _____
ADDRESS _____
[ ] INDIVIDUAL [ ] SMALL BUSINESS CONCERN [ ] NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of payment, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME AND TITLE OF PERSON SIGNING Thomas R. Coolidge, Chairman of the Board and C.E.O.
ADDRESS OF PERSON SIGNING 173 Beebe Hill Rd., Falls Village, CT 06031

SIGNATURE Thomas R. Coolidge DATE 4/19/99

Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)	Attorney's Docket Number P03986US2

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorneys and/or agents to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. *(List name and registration number)* Donald H. Zarley, Reg. #18,543; Bruce W. McKee, Reg. #19,651; Dennis L. Thomte, Reg. #22,497; Michael G. Voorhees, Reg. #25,715; Edmund J. Sease, Reg. #24,741; Mark D. Hansing, Reg. #30,643; Kirk M. Hartung, Reg. #31,021; Daniel J. Cosgrove, Reg. #36,770; Michael R. Crabb, Reg. #37,298; Heidi Sease Nebel, Reg #37,719; Wendy K. Marsh, Reg.# 39,705; Jeffrey D. Harty, Registration No. 40,639; James A. Napier, Registration No. 42,025; Mark Ziegelbein, Registration No. 43,307; Timothy J. Zarley, Registration No. P-45,253; and Patty L. Ades, Registration No. P-44,496.

Send correspondence to: Attorney Name EDMUND J. SEASE ZARLEY, MCKEE, THOMTE, VOORHEES & SEASE 801 Grand Avenue, Suite 3200 Des Moines, Iowa 50309-2721	**Direct Telephone Calls to:** *(Name and telephone number)* Attorney Name EDMUND J. SEASE 515-288-3667

2	FULL NAME OF INVENTOR	FAMILY NAME GÖKE	FIRST GIVEN NAME Burkhard	SECOND GIVEN NAME .
0	RESIDENCE & CITIZENSHIP	CITY Marburg	STATE OR FOREIGN COUNTRY Germany DEX	COUNTRY OF CITIZENSHIP DE
1	POST OFFICE ADDRESS	POST OFFICE ADDRESS Mariborer Str. 22	CITY Marburg	STATE & ZIP CODE/COUNTRY Germany 35037
2	FULL NAME OF INVENTOR	FAMILY NAME BYRNE	FIRST GIVEN NAME Maria	SECOND GIVEN NAME
0	RESIDENCE & CITIZENSHIP	CITY Munster-Hiltrup	STATE OR FOREIGN COUNTRY Germany DEX	COUNTRY OF CITIZENSHIP DE
2	POST OFFICE ADDRESS	POST OFFICE ADDRESS Am Hagen 21	CITY Munster-Hiltrup	STATE & ZIP CODE/COUNTRY Germany D-48165
2	FULL NAME OF INVENTOR	FAMILY NAME COOLIDGE	FIRST GIVEN NAME Thomas	SECOND GIVEN NAME R.
0	RESIDENCE & CITIZENSHIP	CITY Lincoln	STATE OR FOREIGN COUNTRY Nebraska NE	COUNTRY OF CITIZENSHIP US
3	POST OFFICE ADDRESS	POST OFFICE ADDRESS 3820 NW 46th Street	CITY Lincoln	STATE & ZIP CODE/COUNTRY Nebraska 68524

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)	Attorney's Docket Number P03986US2
SIGNATURE OF INVENTOR 201 Burkhard Göke B. G.	SIGNATURE OF INVENTOR 202 Maria Byrne
DATE 2-27-2001	DATE
SIGNATURE OF INVENTOR 203 Thomas R. Coolidge	SIGNATURE OF INVENTOR 204
DATE	DATE

End of Combined Declaration for Patent Application and Power of Attorney

Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)	Attorney's Docket Number P03986US2

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Send correspondence to:
Attorney Name

EDMUND J. SEASE
ZARLEY, MCKEE, THOMTE, VOORHEES & SEASE
801 Grand Avenue, Suite 3200
Des Moines, Iowa 50309-2721

Direct Telephone Calls to:
(*Name and telephone number*)
Attorney Name
EDMUND J. SEASE
515-288-3667

2	FULL NAME OF INVENTOR	FAMILY NAME GÖKE	FIRST GIVEN NAME Burkhard	SECOND GIVEN NAME
0	RESIDENCE & CITIZENSHIP	CITY Marburg	STATE OR FOREIGN COUNTRY Germany	COUNTRY OF CITIZENSHIP DE
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2	FULL NAME OF INVENTOR	FAMILY NAME BYRNE	FIRST GIVEN NAME Maria	SECOND GIVEN NAME
0	RESIDENCE & CITIZENSHIP	CITY Munster-Hiltrup	STATE OR FOREIGN COUNTRY Germany	COUNTRY OF CITIZENSHIP ~~DE~~ IRELAND
2	POST OFFICE ADDRESS	POST OFFICE ADDRESS Am Hagen 21	CITY Munster-Hiltrup	STATE & ZIP CODE/COUNTRY Germany D-48165
2	FULL NAME OF INVENTOR	FAMILY NAME COOLIDGE	FIRST GIVEN NAME Thomas	SECOND GIVEN NAME R.
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Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)	Attorney's Docket Number P03986US2

SIGNATURE OF INVENTOR 201 Burkhard Göke 2-0	SIGNATURE OF INVENTOR 202 Maria Byrne M. Byrne
DATE	DATE 23.02.01
SIGNATURE OF INVENTOR 203 Thomas R. Coolidge	SIGNATURE OF INVENTOR 204
DATE	DATE

End of Combined Declaration for Patent Application and Power of Attorney

Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Application)	Attorney's Docket Number P03986US2

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Send correspondence to: EDMUND J. SEASE ZARLEY, MCKEE, THOMTE, VOORHEES & SEASE 801 Grand Avenue, Suite 3200 Des Moines, Iowa 50309-2721	Direct Telephone Calls to: (*Name and telephone number*) EDMUND J. SEASE 515-288-3667

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Combined Declaration for Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Application)	Attorney's Docket Number P03986US2

SIGNATURE OF INVENTOR 201	SIGNATURE OF INVENTOR 202
Burkhard Göke	Maria Byrne
DATE	DATE
SIGNATURE OF INVENTOR 203 Thomas R. Coolidge *Thomas R. Coolidge*	SIGNATURE OF INVENTOR 204
DATE 2/21/01	DATE

End of Combined Declaration for Patent Application and Power of Attorney